



Contents lists available at ScienceDirect

Scientia Horticulturae

journal homepage: www.elsevier.com/locate/scihorti



Review

Fungus resistant grape varieties as a suitable alternative for organic wine production: Benefits, limits, and challenges

Karine Pedneault^{a,*}, Caroline Provost^{b,*}

^a Centre de développement bioalimentaire du Québec, 1642 rue de la Ferme, La Pocatière, Québec, Canada

^b Centre de recherche agroalimentaire de Mirabel, 9850 Belle-Rivière, Mirabel, Québec, Canada

ARTICLE INFO

Article history:

Received 5 November 2015
Received in revised form 8 March 2016
Accepted 14 March 2016
Available online xxx

Keywords:

Interspecific hybrid
PIWI grape
Organic viticulture
Breeding
Phenolic compounds
Volatile compounds
Resistance
Powdery mildew
Downy mildew
Botrytis

ABSTRACT

Areas dedicated to organic wine production have significantly increased over the last few years. The vast majority of organic wine is made from *Vitis vinifera* varieties that are highly susceptible to fungal diseases and pests, making organic management difficult for growers. Depending on the growing area, 20–70% of organic growers declare issues with fungal diseases in Europe. Recently, fungus-resistant grape (FRG) varieties have been recommended as the most suitable choice in organic viticulture, especially in areas where disease pressure necessitates high rates of fungicides. FRG varieties could contribute to improved disease management in organic as well as conventional viticulture, reduce production costs and decrease copper accumulation in soils. Recently, many FRG varieties presenting advantageous agronomic attributes and enological characteristics have been developed in North America and Europe for conventional and sustainable farming. In this review, we present an overview of the benefits and limits associated with FRG varieties in addition to the current knowledge regarding berry and wine composition, canopy management, and winemaking challenges and practices.

© 2016 Elsevier B.V. All rights reserved.

Contents

1. Introduction	00
2. Benefits and limits of FRG in organic viticulture	00
2.1. Breeding for disease resistance and quality	00
2.2. Economical and agricultural benefits of disease resistance	00
2.3. Yield	00
2.4. Marketing and wine quality	00
3. Growing FRG varieties for organic wine production	00
3.1. Berry and juice composition	00
3.1.1. Juice	00
3.1.2. Berries	00
3.2. Impacts of canopy management	00
3.2.1. Managing yield and berry quality	00
3.2.2. Impact on wine	00
3.3. Challenges in FRG wine production	00
3.3.1. Juice extraction and methanol	00

Abbreviations: FRG, fungus resistant grape; M3GE, malvidin-3-glucoside equivalents; MDGE, malvidin-3,5-diglucoside equivalents; DW, dry weight; FW, fresh weight; LC-MS/MS, liquid chromatography coupled to mass spectrometry; UPLC-MS, ultraperformance liquid chromatography coupled to mass spectrometry; HPLC-DAD, high performance liquid chromatography coupled to diode-array detector; CE, catechin equivalent; B2E, procyanidin B2 equivalent; RE, resveratrol equivalent; TA, titratable acidity; TSS, total soluble solids; YAN, yeast assimilable nitrogen.

* Corresponding authors.

E-mail addresses: karine.pedneault@cdbq.net, karine.pedneault.1@ulaval.ca, karinepedneault@gmail.com (K. Pedneault), cprovost@cram-mirabel.com (C. Provost).

<http://dx.doi.org/10.1016/j.scienta.2016.03.016>

0304-4238/© 2016 Elsevier B.V. All rights reserved.

Please cite this article in press as: Pedneault, K., Provost, C., Fungus resistant grape varieties as a suitable alternative for organic wine production: Benefits, limits, and challenges. *Sci. Hortic.* (2016), <http://dx.doi.org/10.1016/j.scienta.2016.03.016>

3.3.2.	Biogenic amines	00
3.3.3.	Tannins and color	00
3.3.4.	Aroma	00
3.4.	Improving FRG wine quality	00
4.	Conclusion	00
	Acknowledgements	00
	References	00

1. Introduction

Organic wine production considerably increased over the last ten years and accounts for nearly 5% of total production of today's wine market (Bonn et al., 2015; Willer and Lernoud, 2015). Reasons driving consumers toward organic wine varies from health concerns to environmental awareness and interest for *terroir*, and the overall perception that organic wines are of higher value than conventional ones (Bonn et al., 2015).

Organic wines are expected to be free of synthetic pesticides, fertilizers or other synthetic inputs that could pose a higher risk than conventional farming to human health or the environment (Mann et al., 2010; Bonn et al., 2015). However, the vast majority of organic wine is made from *Vitis vinifera* varieties highly susceptible to fungal diseases and pests, making organic management difficult for growers (Wiedemann-Merdinoglu and Hoffmann, 2010). Depending on the country origin, 20–70% of organic growers declare issues with botrytis and powdery mildew in Europe (Collective, 2008).

In contrast with conventional viticulture that integrates a wide range of synthetic pesticides in pest management programs, organic viticulture mostly relies on sulfur- and copper-based fungicides such as the Bordeaux mixture to control major diseases like downy and powdery mildews, as well as a wide range of other diseases and insect pests (Provenzano et al., 2010). Copper-based formulations have been used for more than a hundred years in European vineyards (Flores-Vélez et al., 1996), but copper has a low mobility in soil and was later found to accumulate to levels that could threaten the environment (Komárek et al., 2010). Indeed, copper concentration is typically much higher in vineyard soils (20–665 mg/kg) compared to arable land (5–30 mg/kg) (Besnard et al., 2001; Komárek et al., 2010). Such accumulation is likely to occur in perennial crops like grapevines because copper fungicides are continuously sprayed on the same land (Komárek et al., 2010). Correspondingly, vineyards located in wet areas show higher copper concentration than those from dry areas, which suffer less pressure from diseases (Komárek et al., 2010). Copper concentration reached 93 mg/kg in the 0–20 cm soil layer of a Mediterranean organic vineyard with a dry climate, whereas values between 200 and 297 mg/kg have been reported in conventional vineyards located in wet areas from Northern Italy (Provenzano et al., 2010).

The European Union recently established premium goals to reduce pesticides, particularly copper, in viticulture (Rousseau et al., 2013). One of the strategies is to shift from a treatment-oriented approach to a disease prevention approach by the development of fungus-resistant varieties (Rousseau et al., 2013). Fungus-resistant grapes (FRG) result from interspecific cross-breeding between the Mediterranean species *V. vinifera* and North American and Asian *Vitis spp.* such as *V. riparia*, *V. amurensis* and *V. rupestris* that carries high resistance to fungal diseases, including powdery and downy mildews, and grey rot. The firsts FRG varieties, issued from traditional breeding, carried a significant percentage of non-*V. vinifera* species in their genetic and were therefore considered as “interspecific hybrids” (Sivčev et al., 2010). Recently, marker-assisted selection combined with multiple back-crossing with *V. vinifera* varieties allowed the development of FRG

carrying both disease-resistance genes and a significant percentage (more than 85%) of *V. vinifera* in their pedigree; those are generally referred to as “PIWI” (from German: *Pilzwiderstandsfähige*, “disease resistant”) and are accepted as *V. vinifera* varieties in European catalogues (Sivčev et al., 2010). In some cases though, “PIWI” indistinctly refers to both interspecific hybrids and newer “disease-resistant *V. vinifera*” varieties (Collective, 2008; Siegfried and Temperli, 2008).

Optimal variety selection is a key factor for successful implementation of organic grape production (Fragoulis et al., 2009; Sivčev et al., 2010). Recently, FRG varieties have been recommended as the most suitable choice for organic viticulture (Pavloušek, 2010; Sivčev et al., 2010; Becker, 2013). Resistance to major diseases such as powdery mildew significantly reduces the need for pesticides and thus represents a major advantage in organic farming, especially in humid areas such as Bordeaux (Galbrun, 2008; Sivčev et al., 2010; Wiedemann-Merdinoglu and Hoffmann, 2010; Fuller et al., 2014; Weigle and Carroll, 2015).

Superficies devoted to organic wine production from FRG varieties are not well documented. In Germany, FRG occupied 7.9% of organic vineyard surface areas in 2003 but projections were that 40% of the new plantings planned from 2010 to 2015 would be FRG cultivars (Sloan et al., 2010). In contrast, both old and recent FRG varieties have spread extensively over the last 30 years in areas presenting challenging growing conditions such as wet summers and cold winters with most of these varieties grown using conventional management practices (Table 1). Very little research has been done to compare vineyard practices and pesticide use for FRG varieties grown under conventional vs. organic management. In addition, the large genetic pool found in FRG varieties makes them highly variable in terms of viticulture (e.g., vigor, canopy management, berry ripening), berry composition (e.g., sugars, acids, phenolic compounds) and winemaking (e.g., aroma). In order to obtain a comprehensive understanding of FRG varieties with regards to organic grape production, this review will present the benefits and limits of FRG for organic wine production, present their agronomic and oenological characteristics, and discuss challenges related to their use in wine production.

2. Benefits and limits of FRG in organic viticulture

2.1. Breeding for disease resistance and quality

In phytopathology, “resistance” refers to the capacity of the plant to defend itself against pathogens (Prell and Day, 2001). Interspecific hybridization of grapevines began in the 19th Century and was initially aimed at introducing pest and disease resistance in offspring (Galet 1999; Avenard et al., 2003; Reynolds, 2015). Many FRG developed at that time carried undesirable “foxy” flavors in their wine, a default that was later attributed to the presence of *V. labrusca* in the breeding process, resulting in the near elimination of this species in recent breeding (Hemstad and Luby 2000; Sun et al., 2011a). Later, several breeding programs implemented worldwide led to the development of varieties showing different characteristics such as cold-hardiness, short/long growing season, and pest resistance (detailed review by Reynolds, 2015).

Table 1
Estimated growing area for significant FRG varieties grown in Europe, North America, and Asia.

Color	Variety	Estimated growing area (ha)	Countries	Ref.
red	Chambourcin	200–1000	France, Italy, Switzerland	1 ^a
	Baco Noir	200–1000	Canada, United States	2–5
	Frontenac	200–1000	Canada, China, United States	2–4, 6
	Maréchal Foch	200–1000	Canada, United States, France	2–5
	Marquette	200–1000	Canada, United States	2–4
	Regent	>1000	Austria, Belgium, Bulgaria, Czech Republic, Denmark, England, Germany, Hungary, Italy, Netherlands, Sweden, Switzerland	1
white	Baco Blanc	>1000	France	7
	Bianca	>1500	Austria, Belgium, Czech Republic, Denmark, Hungary, Italy, Moldova, Netherlands, Romania, Serbia, Switzerland	1,7
	Cabernet Blanc	200–1000	Austria, Germany, Switzerland	1
	Frontenac Gris	200–1000	Canada, United States	2–4
	Kunleány	>1500	Hungary	1,7
	La Crescent	200–1000	Canada, United States	3, 4
	Vidal	>1000	Canada	2–5
	Villard blanc	200–1000	France	7
	Zala Gyöngye	>2500	Hungary	7

^a (1) Rousseau et al., 2013; (2) Association des vignerons du Québec, 2013 (Personal communication); (3) Tuck and Gartner, 2013; (4) Mount Kobau Wine Services, 2014; (5) Grape Growers of Ontario, 2015; (6) Li et al., 2015; (7) Eurostat, 1999–2009.

The recent sequencing of the *Vitis* sp. nuclear genome significantly contributed to the implementation of molecular breeding strategies in grapevines (see the review by Di Gaspero and Foria, 2015). These tools assist breeders in the selection of parents or of the most promising offspring (Di Gaspero and Foria, 2015). So far, many markers related to desirable agronomic and enological characteristics (e.g., fruit ripening time, berry size, color, acidity, aroma) have been identified in grapevine, including a number of markers for disease resistance (Emanuelli et al., 2011; Merdinoglu and Caranta, 2013; Di Gaspero and Foria, 2015; Zini et al., 2015). Genes related to disease resistance contribute to the activation of pathogen-specific defense mechanisms (PSDM) that induce a hypersensitive response in plant upon the detection of specific pathogen-encoded Avrulence proteins (Verhagen et al., 2010; Wu et al., 2014). PSDM are among the most effective defenses of plants against pathogens (Wu et al., 2014). The main genetic markers related to PSDM in *Vitis* sp. include markers for downy mildew (*rpv1*, *rpv3*, *rpv10*, *rpv12*) and for powdery mildew (*ren1*, *ren3*, *run1*) (Zini et al., 2015; Di Gaspero et al., 2012).

So far, downy and powdery mildews, and botrytis have been primarily targeted for the development of FRG varieties and several cultivars bearing resistance to these diseases have been developed over the last twenty years (Rousseau et al., 2013). Nevertheless, certain pathogens have been able to bypass the resistance mechanisms of certain FRG in field trials (Di Gaspero and Foria, 2015; Merdinoglu and Caranta, 2013). Recently, the pyramidization of resistance haplotypes from different grape species into new FRG varieties prevented pathogens from bypassing resistance mechanisms, therefore making FRG resistance more durable (Di Gaspero and Foria, 2015; Merdinoglu and Caranta, 2013).

2.2. Economical and agricultural benefits of disease resistance

The benefits of growing FRG cultivars go well upon the sector of organic wine production. Indeed, most *V. vinifera* varieties have low to high susceptibility to fungal diseases that result in significant production costs and economic losses (Fuller et al., 2014). In Italy, the annual cost for controlling downy mildew in conventional vineyard typically ranges from 8 to 16 million Euros per year, depending on disease pressure (Salinari et al., 2006). Under medium disease pressure, 12 treatments per season are necessary for traditional *V. vinifera* varieties grown under conventional management (Rousseau et al., 2013). In a study including 183 FRG

varieties grown in six different European countries, the number of fungicide treatments was reduced by 73% and 82% in organic vineyards with low and medium disease pressure, respectively (Rousseau et al., 2013). In a survey involving 65 German vineyards under organic management, growers reported having to spray FRG varieties 3.8 times per season on average (Becker, 2013).

Estimations are that growing FRG varieties could cut production costs by two in French vineyards (Galbrun, 2008). In California, it has been estimated that powdery mildew resistant varieties could allow cost savings as high as 48 M\$ per year for the production subsets of Crimson Seedless Table grapes, raisin grapes and Central Coast Chardonnay wine grapes (Fuller et al., 2014).

The disease resistance of FRG varies with cultivar's genetic and growing location (Pavloušek et al., 2014). Therefore most FRG varieties show some susceptibility to different pathogens, including downy mildew, powdery mildew, botrytis, black rot and anthracnose (Table 2). In organic management, these diseases are generally controlled using sulfur-based fungicides (i.e., *Myco-San*) (Rousseau et al., 2013; Siegfried and Temperli, 2008). When copper-based formulations are necessary, they are used at a much lower rate than for *V. vinifera* varieties (Van Der Meer and Léville, 2010). In Québec (Canada), garlic powder suspension (marketed as *Buran*) is known among the local agronomists to efficiently prevent powdery mildew in organic FRG.

2.3. Yield

The yield of organic *V. vinifera* grapevines can show 8–16% reduction compared to conventional grapevines (Bayramoglu and Gundogmus, 2008; Guesmi et al., 2012). In contrast, FRG are often more vigorous and may show high productivity (10 000–20 000 T/ha), which could contribute to secure yields in organic production (Reynolds and Vanden Heuvel, 2009; Sun et al., 2011b; Rousseau et al., 2013; Barthe, 2015).

Yields ranging from 7.0 to 11.8 T/ha were reported for FRG grown under conventional management with minimal treatment in Switzerland (Siegfried and Temperli, 2008). In a five years study carried out in New York State, organic Seyval blanc grapevines produced, on average, 30% less crop than vines grown using conventional management (12.7 vs. 19.9 Tons/ha, respectively; Pool, 1995). Such reduction was caused by differences in soil quality (e.g., the organic plots was inferior to that of conventional plots) and by

Table 2
Overview of disease susceptibility, vigor and hardiness/cold tolerance of 41 fungus-resistant varieties.

Variety	Susceptibility to diseases ^{a,b}									Vigor	Hardiness/Cold tolerance (°F)	Ref. ^c
	DM	PM	BOT	BR	AT	DA	CG	EUT				
reds												
Baco noir	+	++	+	+++		+	+++	++	high	–10 to –20	1–7	
Baltica		+		+	+				moderate	–20 to –30	1, 8–10	
Cabernet Carbon	–/+	++	–/+								2, 5	
Cabernet Jura	–	–/+	–/+								2, 5	
Chambourcin	+	+	+++	+++		+	++		moderate/high	–5 to –15	1–6, 11	
Chancellor	+++	++	+	+	+	+++	++	+	moderate	–10 to –20	1–6, 12, 13	
Chelois	+	+++	+	+		+++	++	++		–10 to –20	6	
Concord	+	++	+	+++		+++	+	+++	high	–15 to –25	3, 4, 6	
De Chaunac	++	++	+	+	+	++	++	+++	moderate/high	–15 to –25	1–5	
Frontenac	–/+	++	++	++	+	++			high	–20 to –30	1, 3, 4, 8–10	
Léon Millot	+	++	+	+		+	++	++	high	–15 to –25	1–7, 11	
Lucie Kuhlmann	+	++			+	+			moderate		1, 12, 13	
Maréchal Foch	+	++	+	++		+	+++	+++	moderate	–15 to –25	1–7, 11, 14	
Marquette	–	+	+	+	+++				high	–20 to –30	1, 8–10, 14	
Petite Perle	+	+	+	+	+				moderate	–20 to –30	1, 8, 9, 14	
Regent	+	+	+			+				–5 to –15	2, 5, 7, 15	
Sabrevois	+	+		+	+				moderate	–20 to –30	1, 10, 13, 14	
Skandia	+	+	+	+	++				moderate	–20 to –30	1, 8, 9, 14	
St. Croix	++	++	++	+	+				moderate/high	–20 to –30	1, 3, 6, 8–10, 12–14	
whites												
Aurore	++	++	++	+++		+	++	+++	high	–10 to –20	3–7	
Bianca	+	+	–/+			++				–5 to –15	2, 5, 7, 16	
Bronner	–/+	+	+								5, 2, 11	
Cabernet blanc	++	++	–/+							–5 to –15	2, 5	
Frontenac blanc	–/+	+		++	++				high	–20 to –30	1, 8–10, 14	
Frontenac gris	–/+	+		++	+				high	–20 to –30	1, 8–10, 14	
Geisenheim	+	+++	+		+				moderate		1	
Helios	++	+	+								2, 5, 11	
Hibernal	++	+	+	+		+			moderate		1, 5, 12, 13	
Johanniter	++	–/+	++								2, 5, 11	
Kunleány	++	+++	–								5, 16	
La Crescent	+	+		++	++				high	–20 to –30	1, 3, 8–10, 14	
Louise Swenson	+	+	+	+	++				low	–20 to –30	1, 8–10, 14	
Osceola Muscat	++	++	+	+	+				high	–15 to –25	1, 8, 9	
Seyval	+	+++	++	++	+	+	++	+	moderate	–10 to –20	1–7, 10, 11, 13, 14	
Solaris	++	–/+	++							–10 to –20	2, 5, 11, 14	
Soleil blanc	+	–/+	++								2, 5, 11	
St. Pepin	+	+	+		+				moderate	–20 to –30	1, 6, 10, 12–14	
Traminette	+	+	+	+	+++	++	++	+	moderate/high	–10 to –20	1, 3, 4, 10	
Vandal-Cliche	+	+	+		+++				moderate/high	–15 to –25	1, 10, 12, 13	
Vidal	+	++	–/+	+	+	+	+++	+	moderate	–5 to –15	1–6, 10, 12, 13	
Villard blanc	–	+		+							5, 15	

^a DM: downy mildew; PM: powdery mildew; BOT: *Botrytis*; BR: black rot; AT: anthracnose; DA: dead arm; CG: crown gall; EUT: *Eutypa*.

^b –/+ = resistant and few susceptible according to several references; – = resistant; + = slightly susceptible; ++ = moderately susceptible; +++ = highly susceptible.

^c (1) Dubé and Turcotte, 2011; (2) Sivčev et al., 2010; (3) Dami et al., 2005a; (5) Rousseau et al., 2013; (6) Reisch et al., 1993; (7) Lisek, 2010; (8) Provost et al., 2012b; (8) Wolf, 2008; (9) Provost et al., 2013; (10) Carisse and Lefebvre, 2011; (11) Tuchschnid et al., 2006; (12) Khanizadeh et al., 2008; (13) Khanizadeh et al., 2005; (14) Plocher and Parke, 2008; (15) Avenard et al., 2003; (15) Daniela et al., 2013; (16) Pavloušek, 2013.

the competition caused by cover crops in the organic plot (Pool, 1995).

2.4. Marketing and wine quality

FRG varieties are nearly completely absent from wine marketing in major wine producing countries, a situation that limits their expansion because they remain unknown to consumers. In a survey of 255 wineries growing FRG varieties (25% of them being under organic management), 40% of respondents pointed the “problem of unknown varieties” as the biggest handicap for the marketing of FRG wines (Becker, 2013). As FRG varieties carry non-*V. vinifera* genes (even at low levels), they may suffer from the perception that interspecific hybrids produce low-quality wines (Fuller et al., 2014). Similarly, organic wine has suffered from a reputation of average quality until recently (Collective, 2008). The need to educate consumers to organic wines made from FRG varieties is therefore significant.

The quality of FRG wines has been a key topic since their development in the 19th Century. Back then, many French-American hybrids were thought to produce wine of “satisfactory commercial quality” and were winning medals in wine competitions (Paul, 1996a). Recent studies showed that the quality of FRG wines is generally rated as equivalent to that of *V. vinifera* (Van Der Meer and Léville, 2010; Pedneault et al., 2012; Rousseau et al., 2013). For example, in a blind tasting carried out on 52 FRG wines from Europe, 62% red FRG varieties (24 wines tasted), including Cabernet Jura (VB 5-02), Cabertin (VB 91-26-17) and the old interspecific hybrid Chambourcin (J. Seyve 26-205), were noted as equivalent or superior to Merlot (reference wine), and 31% of white (28 wines tasted) were classified as equivalent or superior to the reference Chardonnay wine, including the interspecific varieties Gf. GA.47-42 (Bacchus Weiss X Seyval blanc), Saphira (Gm 7815-1), and Solaris (Fr 240-75) (Rousseau et al., 2013).

A consumer study conducted in Switzerland concluded that 70–90% of consumers noted Solaris and Maréchal Foch wines as equivalent to *V. vinifera* Riesling and Zweigelt wines

(used as reference wines), respectively, and 23–30% of consumers judged the FRG wines as “clearly superior” to the reference *V. vinifera* wines (Van Der Meer and Léville, 2010). A consumer survey comparing 21 red FRG wines produced in Eastern Canada to three imported *V. vinifera* wines described as major selling products in this area showed that 76% of FRG wines were judged as equivalent or superior to the reference wines (Pedneault et al., 2012). Most FRG wines were blends that included Marechal Foch or Frontenac, with other locally grown FRG varieties (Pedneault et al., 2012).

3. Growing FRG varieties for organic wine production

The chemical composition of FRG is highly variable from one variety to another, which is attributable to their large genetic pool. This entails the need to optimize the relationship between varieties and growing conditions. Knowledge of berry chemical composition and of the impact of canopy management practices is essential to achieve that goal. Studies cited in the following Sections (3.1 and 3.2) were generally conducted under conventional management and in northern areas; this is where most FRG have been grown and studied. To our knowledge, no studies are available yet on canopy management practices for European PIWI varieties. Studies comparing organic to conventional management in *V. vinifera* varieties have been reviewed by Provost and Pedneault (2016) and show that organic management has generally a limited impact on berry and wine composition, and wine quality.

3.1. Berry and juice composition

3.1.1. Juice

An overview of juice characteristics (e.g., TSS, pH, TA) of FRG grown in different locations is presented in Table 3. Certain FRG varieties present high pH and high TA, which causes issues with microbial spoilage and color stability in wines (Morris et al., 1984a). YAN level also varies largely among FRG varieties. The analysis of 30 FRG varieties across Midwestern and Eastern United States showed YAN concentrations ranging between 89 and 938 mg/L (Stewart and Butzke, 2012). High YAN values (≥ 250 mg/L) are common in *V. riparia*-based FRG such as Frontenac (Mansfield, 2015a; Slegers et al., 2015; Stewart, 2013). In addition, significant variations are observed between years, growing areas and cultural practices (Stewart, 2013).

3.1.2. Berries

Red FRG berries typically show anthocyanin concentrations ranging from 0.5 to 1.5 mg/g M3 G eq. FW in berries with low tannin concentration ranging from 0.07 to 0.95 mg/g berry in seed, and from 0.03 and 0.79 mg/g berry in skin (Tables 4 and 5). Conversely, tannin levels in *V. vinifera* cv. Pinot noir reach 1.2 mg/g berry in seed and 0.56 mg/g berry in skin (Springer and Sacks, 2014). The profile of other phenolic compounds such as flavonols and phenolic acid derivatives varies significantly from one FRG variety to another (Tables 6 and 7). In red European PIWI varieties such as Cabernet Cortis and Regent, isorhamnetin-3-*o*-rutinoside is the major flavonol whereas quercetin-3-*o*-glucoside is the major flavonol in Baco noir and Lucy Khulmann (Ratnasooriya et al., 2011; Ehrhardt et al., 2014). Caftaric acid is the major hydroxycinnamic derivative in most FRG varieties (Zhu et al., 2012; Manns et al., 2013; Ehrhardt et al., 2014).

Stilbenes such as resveratrol are known to be involved in plant defense mechanism against fungal infections (Ehrhardt et al., 2014). *trans*-Resveratrol and its glucoside are the main stilbenes reported in berries of French hybrid FRG varieties grown in Canada, whereas high levels of *trans*- and *cis*-piceid, pallitol and astringin have been reported in PIWI varieties from Italy and Germany (Table 8).

3.2. Impacts of canopy management

3.2.1. Managing yield and berry quality

Controlling yield may contribute to enhance berry and wine quality, especially when the growing season is unfavorable to optimal berry ripening (Berkey et al., 2011). Studies show that either cluster or shoot thinning contribute to increased TSS and pH in FRG berries, but results vary from one year to another and, in many trials, harvest date had a higher impact than yield management (Tables 9 and 10). For example, 36% yield reduction using cluster thinning negatively impacted increased the level of C₆ compounds in juice of Seyval blanc berries during a hot growing season in Quebec (Canada) (Pedneault et al., 2015). Reducing yield significantly increased skin softness in Seyval blanc berries, suggesting that it may impact physical barriers protecting berries from fungal infections (Barthe, 2015).

Geneva Double Curtain (GDC) training contributed to increase yield but conversely reduced TSS and pH in a season-dependent manner in Chancellor (Reynolds et al., 1995). GDC and Umbrella-Kniffin (UK) increased yield in Frontenac and Marquette, respectively, but as UK increased TA and lowered TSS and pH in Marquette, GDC increased TSS and pH, and lowered TA in Frontenac when compared to other training systems (Bavougian et al., 2012; Martinson and Particka, 2013). Both Vertical Shoot Positioning (VSP) and High-Cordon (HC) increased the level of free volatile terpenes in Traminette juice compared to GDC, Smart-Dyson (SD) and Scott-Henry (SH) (Ji and Dami, 2008).

3.2.2. Impact on wine

Shoot thinning significantly decreased yield and the level of C₆ alcohols in Maréchal Foch wines, but did not impacted wine sensory perception (Sun et al., 2011b). In contrast, taste and flavors of wines derived from un-thinned control vines were preferred over those derived from cluster-thinned vines in Vandal-Cliche (Pedneault et al., 2015).

Chancellor wines issues from the GDC scored higher on berry flavor than those issued from the Y-trellis system, whereas wines issues from Hudson River Umbrella (HRU) scored higher on color and lower on earthy note intensities (Tables 9 and 10; Reynolds et al., 2004). GDC increased yield and decrease TSS and pH in Seyval blanc, but GDC wines rated higher on melon notes intensity, and lower on earthy and vegetal flavors and astringency when compared to wines from 6-arms Kniffin (6AK) and Y-trellis systems (Reynolds et al., 2004).

In Traminette, increasing cluster sunlight exposition improved wine color and sensory ratings for linalool, rose and spice aromas (Bordelon et al., 2008; Skinkis et al., 2010). Increasing cluster exposition had little impact on aroma intensity in Seyval blanc wines, but wines from exposed clusters were rated as superior (Reynolds et al., 1986).

3.3. Challenges in FRG wine production

Recent findings (Manns et al., 2013; Slegers et al., 2015; Springer and Sack, 2015) show that many aspects of current enology knowledge may have limited application with FRG varieties, because most of them present particular biochemical characteristics. Therefore, particular winemaking practices should be developed for these grapes.

3.3.1. Juice extraction and methanol

Many FRG varieties contain high levels of pectin that necessitate the use of enzymes to increase juice yield at pressing. High pectin levels have been thought to increase methanol concentration in FRG wine (Lee et al., 1975). However, surveys showed that the level of methanol in FRG wines ranges between 20 and 197 mg/L, which

Table 3
Overview of TSS, TA, pH and YAN levels in the juice of 41 fungus-resistant grape varieties from different growing areas.

Variety	Localization		TSS (Brix)	TA			pH	YAN (mg/L)	Ref.
	Country	Area		Value	Units	Eq.			
<i>reds</i>									
Baco noir	Canada	Nova Scotia	18.5	0.82	%	tartaric acid	2.66		1 ^a
Baltica	Canada	Québec	24.9	6.96	g/L	tartaric acid	3.05	179	2
Cabernet Carbon	Switzerland	Stäfa	22.5–22.9	7.8–8.3	g/L	n.a.	3.0–3.1		3
	Switzerland	Wädenswill	21.0	10.7	g/L	n.a.	3.0		4
Cabernet Cortis	Switzerland	Wädenswill	22.3	10.6	g/L	n.a.	3.1		4
Cabernet Jura	Switzerland	Wädenswill	21.2–24.0	8.1–10.2	g/L	n.a.	3.0–3.4		3
Chambourcin	China	Beijing	20.2	4.2	g/L	H ₂ SO ₄	n.a.		5
	Switzerland	Wädenswill	19.0–21.6	12–12.4	g/L	n.a.	n.a.		4
Chancellor	United States	Michigan	20.6–21.8	7.2–7.9	g/L	n.a.	3.23–3.34		6
	United States	Ohio	20.1–21.3	10.7–11.2	g/L	n.a.	3.20–3.27		7
	Canada	British Columbia	18.5–23.5	12.2–16.0	g/L	n.a.	3.12–3.37		8
Chelois	United States	Arkansas	14.8–17.1	0.74–0.93	%	tartaric acid	3.47–3.79		9
	United States	Arkansas	14.8–18.2	0.68–1.07	%	tartaric acid	3.61–3.80		9
Corot noir	United States	New York	15.0–17.8	7.5–11.6	g/L	n.a.	3.34–3.75	213	10, 11
Frontenac	Canada	Québec	21.5–25.3	10.1–17.5	g/L	tartaric acid	3.10–3.35	191–403	2,12–14
	United States	Iowa	21.4	10.4	g/L	tartaric acid	3.39		15
	United States	Minnesota	22.9–26.4	14.9–18.5	g/L	tartaric acid	2.86–3.12		16
	United States	Nebraska	19.7–22.5	14.6–20.9	g/L	n.a.	2.92–3.12		17
Léon Millot	Canada	Nova Scotia	20.6	0.81	%	tartaric acid	3.13		1
	Switzerland	Wädenswill	21.6	7.24	g/L	n.a.	3.46		18
Lucy Khulman	Canada	Nova Scotia	21.1	0.76	%	tartaric acid	3.11		1
Maréchal Foch	Canada	Nova Scotia	23.1	0.94	%	tartaric acid	3.06		1
	Canada	Québec	21.6	7.24–10.3	g/L	tartaric acid	3.18	95–108	12,14
	Switzerland	Wädenswill	21.6	9.7	g/L	n.a.	3.2		4
	United States	Minnesota	24.6–26.1	8.7–11.8	g/L	tartaric acid	2.99–3.05		16
Marquette	United States	New York	20.1–25.1	8.67–11.4	g/L	n.a.	3.10–3.70	119	10, 19
	Canada	Québec	23.7–28.2	8.25–13.1	g/L	tartaric acid	3.09–3.50	210–342	2, 12–14
	United States	Iowa	22.7	8.2	g/L	tartaric acid	3.39		15
Noiret	United States	Minnesota	26.2–30.5	11.5–12.0	g/L	tartaric acid	2.84–3.05		16
	United States	New York	21.9	9.37	g/L	n.a.	3.09	329	10
	United States	New York	19.3	7.2	g/L	tartaric acid	3.45	164	20
Petite Perle	Canada	Québec	22.9	7.38	g/L	tartaric acid	3.33	299	2
Regent	Germany	n.d.	20.1–20.3	8.0–8.3	g/L	n.a.	–		21
	Switzerland	Stäfa	20.3–22.0	6.1–8.4	g/L	n.a.	3.3–3.7		3
Sabrevois	Canada	Québec	18.6–19.2	13.4	g/L	tartaric acid	3.16	213–221	12–14
Skandia	Canada	Québec	29	6.84	g/L	tartaric acid	3.69	351	2
St. Croix	Canada	Québec	19.4–22.7	5.14–9.0	g/L	tartaric acid	3.21–3.39	129–167	2, 12, 14
	United States	Iowa	16.8	6.3	g/L	tartaric acid	3.73		15
	United States	Minnesota	20.9–24.1	3.87–4.96	g/L	tartaric acid	3.08–3.50		16
Villard noir	United States	Arkansas	15.1–17.0	0.75–1.00	%	tartaric acid	3.63–3.73		9
<i>whites</i>									
Adalmiina	Canada	Québec	19.4	7.56	g/L	tartaric acid	2.99	183	2
Aurore	United States	Arkansas	17.0–19.0	0.64–0.80	%	tartaric acid	3.68–3.85		9
Bronner	Switzerland	Wädenswill	19.0–22.5	7.3–9.6	g/L	n.a.	3.1		4
Cal 6-04	Switzerland	Bevaix	21.0–24.5	10.2–12.4	g/L	n.a.	2.9–3.0		3
Frontenac blanc	Canada	Québec	26.2	9.77	g/L	tartaric acid	3.27	372	2
Frontenac gris	Canada	Québec	26.8	9.91	g/L	tartaric acid	3.23	369	2
	United States	Minnesota	23.4–27.3	14.3–16.1	g/L	tartaric acid	2.93–3.08		16
Helios	Switzerland	Wädenswill	21.6	8.5	g/L	n.a.	3.2		4
Johanniter	Switzerland	Wädenswill	20.5	9	g/L	n.a.	3.2		4
La Crescent	Canada	Québec	22.8	14.2	g/L	tartaric acid	3	130	2
	United States	Minnesota	22.7–25.8	13.2–14.4	g/L	tartaric acid	2.88–2.99		16
	United States	Iowa	21.8	8.9	g/L	tartaric acid	3.36		15
Noah	China	Beijing	19.9	6.9	g/L	H ₂ SO ₄			5
Osceola Muscat	Canada	Québec	23.5	8.72	g/L	tartaric acid	3.08	163	2
Seyval	Canada	Québec	17.7–22.4	8.77–12.3	g/L	tartaric acid	2.86–3.16	94–180	22
	United States	Arkansas	16.9–17.6	0.60–0.89	%	tartaric acid	3.69–3.91		9
Solaris	Switzerland	Stäfa	26.2–26.9	5.7–7.3	g/L	n.a.	3.2–3.5		3
	Switzerland	Wädenswill	24.9–30.4	5.8–9.5	g/L	n.a.	3.3		4
Soleil blanc	Switzerland	Wädenswill	22.9	8.73	g/L	n.a.	3.43		18
St. Pepin	United States	Minnesota	22.0–23.8	8.8–9.8	g/L	tartaric acid	2.94–3.25		16
Traminette	United States	New York	20.7	7.9	g/L	tartaric acid	3.18	95	20
Vandal-Cliche	Canada	Québec	17.3–19.7	11.2–13.4	g/L	tartaric acid	2.85–2.92	99–162	22
Verdelet	United States	Arkansas	16.2–20.1	0.35–0.84	%	tartaric acid	3.63–4.32		9
Vidal blanc	United States	Michigan	18.8–20.2	0.65–1.02	%	tartaric acid	3.28–3.30		23
Villard blanc	China	Beijing	20.1	6.8	g/L	H ₂ SO ₄			5

^a (1) Ratnaooyiya et al., 2011; (2) Provost et al., 2012a; (3) Siegfried and Temperli, 2008; (4) Tuchschnid et al., 2006; (5) Zhu et al., 2012; (6) Miller et al., 1996; (7) Dami et al., 2006; (8) Reynolds et al., 1995; (9) Morris et al., 1984b; (10) Manns et al., 2013; (11) Sun et al., 2012; (12) Slegers et al., 2015; (13) Pedneault et al., 2013b; (14) Pedneault et al., 2013a; (15) Vos, 2014; (16) Haggerty, 2013; (17) Bavougian et al., 2012; (18) Striby, 2006; (19) Sun et al., 2011b; (20) Nisbet et al., 2014; (21) Eibach and Töpfer, 2003; (22) Barthe et al., 2012; (23) Wolpert et al., 1983.

Table 4
Anthocyanin profile and total anthocyanin concentration in berries (skin, whole berry) and wines of fungus-resistant grape varieties.

Variety	Origin	Dp-3-O-glucoside*	Cy-3-O-glucoside	Pt-3-O-glucoside	Pn-3-O-glucoside	Mv-3-O-glucoside	Pl-3-O-glucoside	Dp-3,5-O-diglucoside	Cy-3,5-O-diglucoside	Pn-3,5-O-diglucoside	Mv-3,5-O-diglucoside	Pl-3,5-O-diglucoside	Pt-3,5-O-diglucoside	
<i>skin</i>														
Corot noir	NY, USA													
Maréchal Foch	NY, USA													
<i>berry</i>														
Baco Noir	NS, Canada	190	40	150		190								
Cabernet cortis	Italy	66.6	2.93	30.1	4.84	79	nd	–	3.11	21.7	1.95	0.04	–	
Léon Millot	NS, Canada	130	21	100		96								
Lucy Khulman	NS, Canada	200	55	200		220								
Maréchal Foch	NS, Canada	250	42	130		110								
Regent	Germany	408	30	87	57	234	0.72	–	9.84	59.7	0.61	0.22	–	
	Italy	221	24.4	71.3	41.5	203	0.32	–	4.7	60.1	1.3	0.17	–	
<i>wine</i>														
Chambourcin	China OH, USA				4.1	7.3					50.6		6.1	
Corot noir	NY, USA							34.0–49.0	15.7–19.8	25.6–29.5	148–163		44.9–62.3	
Maréchal Foch	NY, USA	14.5–18.0	1.00–1.54	12.0–16.7	0.77–1.05	12.2–17.0								
	NY, USA	12.6–56.3	1.14–2.21	16.1–49.0	1.62–3.39	45.1–89.6		2.88–7.39	1.98–2.39	6.12–7.01	50.5–56.9		4.71–6.66	
Marquette	NY, USA	6.72–14.9	0.78–0.84	9.38–16.6	0.87–1.39	28.9–31.1		12.2–15.0	3.47–3.60	15.3–17.8	128–153		22.4–23.8	
Variety	<i>Pn</i> -3-(6'-acetyl)-O-glucoside	<i>Mv</i> -3-(6'-acetyl)-O-glucoside	<i>Dp</i> -3-(6'- <i>p</i> -coumaroyl)-O-glucoside	<i>Cy</i> -3-(6'- <i>p</i> -coumaroyl)-O-glucoside	<i>Pt</i> -3-(6'- <i>p</i> -coumaroyl)-O-glucoside	<i>Pn</i> -3-(6'- <i>p</i> -coumaroyl)-O-glucoside	<i>Mv</i> -3-(6'- <i>p</i> -coumaroyl)-O-glucoside	Total anthocyanin	Units	Method	Ref.			
<i>skin</i>											0.994–1.26	mg/g FW ^a	a ^d	1 ^e
<i>berry</i>											4.64–8.34	mg/g FW ^a	a	2
	1.5	72.2	40.9	7.15	31.5	4.44	128		mg/100 g DW	b	3			
									mg/kg FW	c	4			
									mg/100 g DW	b	3			
									mg/100 g DW	b	3			
	2.87	15.6	76	60	36	34	91		mg/kg FW	c	4			
	3.12	29.9	164	49.9	69	36.4	313		mg/kg FW	c	4			
<i>wine</i>											84.6	mg/L ^a	b	5
								224–1138	mg/L ^b	d	6			
								671–897	mg/L ^a	a	1			
									mg/L ^c	e	7			
								479–919	mg/L ^a	a	2			
									mg/L ^c	e	7			
									mg/L ^c	e	7			

*Dp: Delphinidin; Cy: Cyanidin; Pt: Petunidin; Pn: Peonidin; Mv: Malvidin; Pl: Pelargonidin.

^a Quantified in malvidin-3-glucoside eq.

^b Quantified in malvidin-3,5-diglucoside eq.

^c Monoglucosides are quantified in malvidin-3-glucoside eq.; diglucosides are quantified in malvidin-3,5-diglucoside eq.

^d (a) Adams-Harbertson protein precipitation assay; (b) LC-MS/MS; (c) UPLC-MS/MS; (d) Spectrophotometry UV-vis at λ_{520nm} ; (e) HPLC-DAD.

^e (1) Sun et al., 2012; (2) Sun et al., 2011b; (3) Ratnaoorya et al., 2011; (4) Ehrhardt et al., 2014; (5) Zhu et al., 2012; (6) Prajitna et al., 2007; (7) Manns et al., 2013.

is slightly higher than *V. vinifera* wines (26–111 mg/L) but significantly lower than the recommended limits of OIV for both reds (≤ 400 mg/L) and whites (≤ 250 mg/L) (Lee et al., 1975; Organisation Internationale de la Vigne et du Vin, 2011).

3.3.2. Biogenic amines

Exogenous nitrogen sources are routinely used to ferment *V. vinifera* varieties. In FRG carrying high YAN levels (see Section 3.1.1), this enrichment may cause an unwanted rise in fermentation temperature and contribute to increase the level of fusel alcohols, undesirable biogenic amines (e.g., histamine) and carcinogenic

Table 5
Flavan-3-ols profile and total tannin concentration in berries (seed, skin, whole berry) and wines of fungus-resistant grape varieties. Data on *Vitis vinifera* (Pinot noir var.) are presented for comparison purposes (from Springer and Sacks, 2014).

Matrix	Variety	Origin	(+)-catechin	(-)-epicatechin	epigallocatechin	(-)-epicatechin 3-O-gallate
<i>seed</i> <i>red</i>	Baco noir	NY, USA				
		ON, Canada	98	106		
	Corot noir	NY, USA				
		NY, USA				
	DeChaunac	ON, Canada	135	78		
		QC, Canada				
	Frontenac	NY, USA				
	Léon Millot	NY, USA				
	Maréchal Foch	ON, Canada	46	42		
		QC, Canada				
	Marquette	QC, Canada				
	Noiret	NY, USA				
	Sabrevois	QC, Canada				
	St. Croix	QC, Canada				
	Vincent	ON, Canada	155	284		
	Pinot noir*	NY, USA				
<i>white</i>	Seyval	ON, Canada	21	23		
<i>skin</i> <i>reds</i>	Baco noir	NY, USA				
	Corot noir	NY, USA				
		NY, USA				
	DeChaunac	NY, USA				
	Frontenac	QC, Canada				
	Léon Millot	NY, USA				
	Maréchal Foch	NY, USA				
		QC, Canada				
		Marquette	QC Canada			
		Noiret	NY, USA			
	Sabrevois	QC Canada				
	St. Croix	QC Canada				
	Pinot noir*	NY, USA				
<i>berry</i> <i>red</i>	Baco Noir	NS, Canada	1700	1100	15	140
	Cabernet cortis	Italy	94.5	101		
	Léon Millot	NS, Canada	860	600	11	99
	Lucy Khulman	NS, Canada	1700	780	11	88
	Maréchal Foch	NS, Canada	2100	990	15	99
	Regent	Germany	176	148		57.4
		Italy	167	115		36.7
<i>white</i>	Johanniter	Italy	84.9	42.1		
	Phoenix	Germany	231	187		121.7
		Italy	64.2	16.5		
	Solaris	Italy	81	111		12.7
<i>wine</i> <i>red</i>	Baco noir	NY, USA				
	Chambourcin	China	1.23	1.23	3.28	
		GA, USA	4.1–9.3	2.6–14.8		
	Corot noir	NY, USA				
		NY, USA	19.3–36.8	16.4–32.8		
		NY, USA				
	Frontenac	QC Canada				
	Maréchal Foch	NY, USA	36–337	11.5–62.1		
		NY, USA				
	Marquette	QC, Canada	13.9–30.0	5.55–14.4		
	NY, USA					
	Sabrevois	QC, Canada				
	St. Croix	QC, Canada				
	Noiret	NY, USA				
	Pinot noir*	NY, USA				

Table 5 (Continued)

Matrix	epigallocatechin gallate	procyanidin B1	procyanidin B2	procyanidin B3	procyanidin B4	procyanidin C1	total tannins	Units	Method	Ref.	
seed red		33	91	41	127	10	0.454	mg/g berry CE	a ^b	1 ^c	
								mg/100 g seed CE or B2E ^a	b	2	
							0.33–0.53	mg/g berry CE	a	3	
							0.917	mg/g berry CE	a	1	
							0.264	mg/g berry CE	a	1	
		4	22	2	12	tr		mg/100 g seed CE or B2E	b	2	
							0.200	mg/g epicatechin eq.	c	4	
							0.594	mg/g berry CE	a	1	
							0.763	mg/g berry CE	a	1	
		14	43	23	61	4		mg/100 g seed CE or B2E	b	2	
							0.377	mg/g epicatechin eq.	c	4	
							0.259	mg/g epicatechin eq.	c	4	
							0.208	mg/g berry CE	a	1	
							0.069	mg/g epicatechin eq.	c	4	
		60	106	27	45	tr	0.170	mg/g epicatechin eq.	c	4	
							mg/100 g seed CE or B2E	b	2		
						1.19	mg/g berry CE	a	1		
white		3	9	2		0		mg/100 g seed CE or B2E	b	2	
skin reds							0.178	mg/g berry CE	a	1	
							0.19–0.31	mg/berry CE	a	3	
							0.34	mg/g berry CE	a	1	
							0.178	mg/g berry CE	a	1	
							0.051	mg/g epicatechin eq.	c	4	
							0.221	mg/g berry CE	a	1	
							0.248	mg/g berry CE	a	1	
							0.030	mg/g epicatechin eq.	c	4	
							0.055	mg/g epicatechin eq.	c	4	
							0.785	mg/g berry CE	a	1	
							0.076	mg/g epicatechin eq.	c	4	
							0.192	mg/g epicatechin eq.	c	4	
							0.559	mg/g berry CE	a	1	
	berry red	8.7							mg/kg DW	d	5
			43.8	106 [#]	28.2	#			mg/kg FW berries	e	6
7.5								mg/kg DW	d	5	
7								mg/kg DW	d	5	
8								mg/kg DW	d	5	
		54.7	64.7 [#]	30.9	#			mg/kg FW berries	e	6	
		65.3	93.7 [#]	45.5	#			mg/kg FW berries	e	6	
white		37.8	50.2 [#]	23	#			mg/kg FW berries	e	6	
		66.8	59.3 [#]	38.4	#			mg/kg FW berries	e	6	
		19.3	22.6 [#]	91.7	#			mg/kg FW berries	e	6	
		3.6	113 [#]	17.9	#			mg/kg FW berries	e	6	
wine red							49	mg/L CE	a	1	
								mg/L CE	d	7	
				2.5–9.6				mg/L CE	f	8	
							42.1–63.6	mg/L CE	a	3	
								mg/L	f	9	
							113	mg/L CE	a	1	
							85.5	mg/L epicatechin eq.	c	4	
							83	mg/L CE	a	1	
								mg/L	f	9	
							120	mg/L epicatechin eq.	c	4	
								mg/L	f	9	
							145	mg/L epicatechin eq.	c	4	
							200	mg/L epicatechin eq.	c	4	
							97.3	mg/L epicatechin eq.	c	4	
							354	mg/L CE	a	1	
						358	mg/L CE	a	1		

* *Vitis vinifera*.

Procyanidins B2 and B4 quantified together.

^a Monomers are quantified as CE; dimers and trimers are quantified as B2E.

^b Analytical method: (a) Adams-Harbertson protein precipitation assay; (b) HPLC-UV-vis; (c) HPLC-Fluorescence; (d) LC-MS/MS; (e) UPLC-MS/MS; (f) HPLC-DAD.

^c (1) Springer and Sacks, 2014; (2) Fuleki and Ricardo da Silva, 1997; (3) Sun et al., 2012; (4) Pedneault et al., 2014; (5) Ratnasooriya et al., 2011; (6) Ehrhardt et al., 2014; (7) Zhu et al., 2012; (8) Auw et al., 1996; (9) Manns et al., 2013.

Table 6

Overview of the flavonol content in berries and wines of some red and white fungus-resistant grape varieties.

Matrix	Variety	Origin	kaempferol-3-o-glucoside	quercetin-3-o-glucoside	quercetin-3-o-glucuronide	isorhamnetin-3-o-rutinoside	quercetin rutin	syringetin-3-glucoside	Units	Method	Ref.	
berry	red	Baco Noir	NS, Canada	20					mg/kg DW	a ^b	1 ^c	
		Cabernet cortis	Italy	3.18	34.1	20.9	50.9		mg/kg FW	b	2	
		Leon Millot	NS, Canada		16				mg/kg DW	a	1	
		Lucy Khulman	NS, Canada		35				mg/kg DW	a	1	
		Marechal Foch	NS, Canada		8				mg/kg DW	a	1	
	white	Regent	Germany	4.04	45.6	60	87.2		mg/kg FW	b	2	
			Italy	nd	8.1	18.9	163.1		mg/kg FW	b	2	
		Johanniter	Italy	5.38	20.2	10.3	nd		mg/kg FW	b	2	
		Phoenix	Germany	4.42	17.2	24.7	nd		mg/kg FW	b	2	
			Italy	nd	16	6.44	nd		mg/kg FW	b	2	
	Solaris	Italy	3.65	32.7	10.9	nd		mg/kg FW	b	2		
wine	red	Chambourcin	China	6.84				5.63	5.69	mg/L ^a	a	3
		Corot noir	NY, USA					ND-0.70	0.65–7.72	mg/L	c	4
		Maréchal Foch	NY, USA					0.54–1.29	0.82–2.97	mg/L	c	4
		Marquette	NY, USA					ND-0.16	2.07–4.68	mg/L	c	4
	white	Noah	China		1.49			2.37	3.1	mg/L ^a	a	3
		Villard blanc	China		ND			3.31	0.2	mg/L ^a	a	3

^a Quantified in mg/L quercetin eq. (QE).^b (a) LC-MS/MS; (b) UPLC-MS/MS; (c) HPLC-DAD.^c (1) Ratnasooriya et al., 2011; (2) Ehrhardt et al., 2014; (3) Zhu et al., 2012; (3) Auw et al., 1996; (4) Manns et al., 2013.**Table 7**

Overview of the hydroxycinnamic and benzoic acids derivatives content in berries and wines of some red and white fungus-resistant grape varieties.

Matrix	Variety	Origin	Hydroxycinnamic acid and derivatives									Benzoic acid derivatives				Units	Method	Ref.				
			caffeic acid	caftaric acid	cinnamic acid	coumaric acid	coutaric acid	caffeic acid ethyl ester	coumaric acid ethyl ester	ellagic acid	fertaric acid	GRP	dihydroxybenzoic acid	gallic acid	protocatechuic acid				ethyl gallate			
berry	red	Cabernet cortis	Italy		54.6			25.7			2.97	3.96			ND				mg/kg FW	a ^c	1 ^d	
		Regent	Germany		36			30			4.78	1.75			0.02				mg/kg FW	a	1	
			Italy		12.7			8.72				1.81	2.12			ND				mg/kg FW	a	1
	white	Johanniter	Italy		17.9			6.44			2.59	4.93			ND					mg/kg FW	a	1
		Phoenix	Germany		26.3			6.49			2.28	1.79			0.03					mg/kg FW	a	1
			Italy		1.02			0.9			1.54	1.57			ND					mg/kg FW	a	1
		Solaris	Italy		15.5			6.93			2.44	1.79			ND					mg/kg FW	a	1
wine	red	Chambourcin	China	2.2				5.76			1.09				7.79	1.63	4.81		mg/L ^a	b	2	
			GA, USA			0.7–15.3		nd-9.3							2.9–17.0				mg/L ^b	c	3	
		Corot noir	NY, USA	4.40–11.50	7.67–30.4	ND-0.25	1.36–14.2	2.09–13.5	0.05–1.17	0.07–2.02		0.54–1.76	0.61–3.68	0.60–1.61		4.64–10.7	0.93–1.30		mg/L	c	4	
		Maréchal Foch	NY, USA	2.79–4.02	8.52–44.1	ND-0.37	3.54–4.82	1.18–10.6	ND-0.61	ND-0.71		1.48–2.65	2.37–3.84	ND-2.09		6.19–12.6	1.41–1.68		mg/L	c	4	
		Marquette	NY, USA	2.98–6.64	26.7–58.8	ND	0.43–2.25	4.79–7.24	0.01–1.17	ND-0.37		1.23–1.33	3.09–6.64	ND-1.47		4.45–8.00	0.57–0.82		mg/L	c	4	

^a Hydroxycinnamic derivatives are quantified in mg/L caffeic acid eq. (CAE); benzoic acid derivatives are quantified in mg/L gallic acid eq. (GAE).^b Quantified in mg/L gallic acid eq. (GAE).^c (a) UPLC-MS/MS; (b) HPLC-ESI-MS/MS; (c) HPLC-DAD.^d (1) Ehrhardt et al., 2014; (2) Zhu et al., 2012; (3) Auw et al., 1996; (4) Manns et al., 2013.

Table 8Overview of the stilbene content in berries and wines of fungus-resistant grape varieties. Data on *Vitis vinifera* wines, from Naugler et al. (2007), are shown for comparison purposes.

Matrix	Variety	Origin	<i>trans</i> -resveratrol	<i>cis</i> -resveratrol	<i>trans</i> -resveratrol glucoside	<i>trans</i> -piceic	<i>cis</i> -piceid	total piceid	pallidol	astringin	Units	Method	Ref.	
whole berry	<i>red</i>													
	Baco Noir	NS, Canada	4.5		9.8						mg/kg DW	a ^b	1 ^c	
	Cabernet cortis	Italy	nd	0.01		0.23	0.41		0.22	0.26	mg/kg FW	b	2	
	Leon Millot	NS, Canada	4.3		9.0						mg/kg DW	a	1	
	Lucy Khulman	NS, Canada	4.3		8.5						mg/kg DW	a	1	
	Maréchal Foch	NS, Canada	4.7		7.8						mg/kg DW	a	1	
	Regent	Germany	1.94	0.17		7.05	11.3		5.62	4.01	mg/kg FW	b	2	
		Italy	nd	0.01		0.21	0.44		0.48	0.28	mg/kg FW	b	2	
	<i>white</i>													
	Johanniter	Italy	nd	0.01		0.08	0.18		0.16	0.13	mg/kg FW	b	2	
	Phoenix	Germany	nd	0.02		0.31	0.57		nd	0.15	mg/kg FW	b	2	
		Italy	nd	nd		0.08	0.16		1.94	0.18	mg/kg FW	b	2	
	Solaris	Italy	nd	nd		0.09	nd		0.26	0.13	mg/kg FW	b	2	
	wine	<i>red</i>												
Baco noir		NS, Canada	0.86	3.68				3.57	0.26	0.16	mg/L RE ^a	c	3	
		NS, Canada	0.90	0.80				3.95	0.14	0.18	mg/L RE	c	3	
Chambourcin		China				tr	nd				mg/L RE	a	4	
		OH, USA	1.4–6.4	0.5–1.9		9.3–29.6					mg/L	d	5	
de Chaunac		NS, Canada	0.65	0.41				7.02	0.36	0.30	mg/L RE	c	3	
Leon Millot		NS, Canada	0.49	0.37				0.96	0.09	0.10	mg/L RE	c	3	
		NS, Canada	0.75	0.38				1.13	0.40	0.17	mg/L RE	c	3	
Leon Millot/Lucie Kuhlman		NS, Canada	1.17	0.80				2.84	0.28	0.22	mg/L RE	c	3	
		NS, Canada	1.92	0.95				2.36	0.38	0.10	mg/L RE	c	3	
Lucie Kuhlman		NS, Canada	0.93	0.68				1.64	0.16	0.16	mg/L RE	c	3	
Maréchal Foch		NS, Canada	0.15	0.74				0.64	0.06	0.04	mg/L RE	c	3	
		NS, Canada	4.55	1.76				2.81	0.315	0.35	mg/L RE	c	3	
		NS, Canada	0.99	0.65				1.34	0.25	0.13	mg/L RE	c	3	
		NS, Canada	1.10	1.0				2.88	0.25	0.30	mg/L RE	c	3	
<i>white</i>														
Noah		China				0.16	nd				mg/L RE	a	4	
Villard blanc		China				0.5	nd				mg/L RE	a	4	
<i>V. vinifera reds</i>														
Zweigelt/Cabernet		NS, Canada	0.56	0.44				1.29	0.11	0.04	mg/L RE	c	3	
Pinot noir/Cabernet	NS, Canada	0.56	0.45				1.56	0.24	0.12	mg/L RE	c	3		
Pinot noir	NS, Canada	0.62	0.58				1.72	0.19	0.06	mg/L RE	c	3		

^a Quantified in mg/L resveratrol eq. (RE).^b (a) HPLC-MS/MS; (b) UPLC-MS/MS; (c) HPLC-DAD.^c (1) Ratnasooriya et al., 2011 ;(2) Ehrhardt et al., 2014; (3) Naugler et al., 2007; (4) Zhu et al., 2012; (5) Prajitna et al., 2007.

Table 9
Impact of cultural practices on grapevine, berries, juices and wine from red varieties.

treatment	variety	# year	main effects observed on			ref.
			grapevine	berry/juice	wine	
cluster thinning	Chambourcin	2–4	yield increased with crop level; thinning favored lignification and reduced bud cold injury	thinning increased TSS, TA, and pH		1 ^a
	Chambourcin	5	thinning reduced yield, increased cluster and berry weight	thinning increased TSS, and pH		2
	DeChaunac	3	same yield and berry weight	no impact on TA, TSS, and pH		3
	DeChaunac	2		thinning increased TSS and total phenols		4
	Marquette	nd	thinning reduced yield	thinning increased TSS and pH		5
	Baco noir	3	pruning reduced yield, cluster weight	pruning increased pH and TSS		6
	Chancellor	3	thinning increase cluster and berry weight	thinning increased TSS, pH, and red color		7
shoot thinning	Concord	nd	thinning increase cluster weight	thinning increased TSS		8
	Frontenac	1	similar cluster weight, reduced berry weight			9
	Frontenac	1	6 shoot/ft had highest cluster number and yield	no impact on TA, TSS, and pH		10
	Maréchal Foch	2	thinning reduced yield, increased berry weight	thinning increased TSS in juice, and berry anthocyanins	thinning decreased the level of C ₆ alcohols (<i>cis</i> -3-hexenol, <i>trans</i> -2-hexenol, 1-hexanol) in wines but impact on wine sensory perception were minor; no impact on wine anthocyanin	11
	Chancellor	5	GDC ^b and YT increased yield, reduced cluster weight and showed optimal level of pruning weight when compared to HRU, 6AK, MWC	When significant, GDC showed TSS, TA and pH in the lowest range, and level of anthocyanin in the highest range		12
training system	Chancellor	4	GDC and 6AK had highest yields; 6AK and YT had highest cluster weight	6AK and YT had the lowest TSS and the highest pH	6AK wines had lowest% ethanol, TA and pH; GDC wines had the highest berry flavour and YT the lowest; HRU wines had the highest color intensity and the lowest intensity in earthy notes no impact on wine anthocyanins	13
	Frontenac	2	GDC increased yield	GDC had higher pH and TSS, and lower TA		14
	Frontenac	nd		VSP had higher total phenol concentration than SD		15
	Marquette	nd	UK increased yield, cluster weight, and clusters/vine	VSP increased TSS and pH; UK had lowest TSS and pH, and higher TA		16
	Marquette	2	TWC had higher yield and cluster weight	TWC had lower TSS; cluster thinning increased TSS in TWC and VSP 18"		17
training system and cluster thinning	St. Croix	2	TWC increased yield and cluster weight; cluster thinning reduced yield	TWC had higher TSS; cluster thinning increased TSS		17

^a (1) Dami et al., 2005b; (2) Dami et al., 2006; (3) Morris et al., 1987; (4) Wood and Looney, 1977; (5) Emling and Sabbatini, 2013; (6) Byrne and Howell, 1978; (7) Morris et al., 2004; (8) Zabadal et al., 2002; (9) Rolfes et al., 2012; (10) Rolfes, 2014; (11) Sun et al., 2011b; (12) Reynolds et al., 1995; (13) Reynolds et al., 2004; (14) Bavougian et al., 2012; (15) Bavougian et al., 2013; (16) Martinson and Particka, 2013; (17) Provost et al., 2012b.

^b 6AK: 6-arm Kniffin; GDC: Geneva double curtain; HRU: Hudson River Umbrella; SD: Smart-Dyson; TWC: Top Wire Cordon; UK: Umbrella Kniffin; VSP: Vertical Shoot Positioning; YT: Y-trellis.

Table 10
Impact of cultural practices on grapevine, berries, juices and wine from white fungus-resistant grape varieties.

treatment	variety	# years	main effects observed on			ref.
			grapevine	berry/juice	wine	
cluster thinning	Seyval	4	yield was similar between treatments; thinning increased berry weight	no impact on TA, TSS, pH		1 ^a
	Seyval	1	thinning increased cluster weight, reduced yield	thinning increased TSS		2
	Vandal-Cliche	2	thinning reduced yield	no impact on TA, TSS, pH		2
	Vidal	2	thinning increased cluster weight	thinning increased TSS/TA ratio		3
leaf removal	Frontenac gris	nd	no impact on berry size	no impact on TSS, TA, pH		4
leaf removal, cluster thinning cluster exposition vs shading	La Crescent	nd	no impact on berry size	leaf removal + no thinning increased TSS		4
	Seyval	2	higher occurrence of bunch rot in shaded clusters	higher TSS and lower TA in exposed clusters	non significant differences between intensity of aroma, perceived acidity and quality of wines, but authors stated that wines from exposed clusters were “superiors”	5
shoot and cluster thinning	Traminette	3		full shaded clusters had lower TSS and PVT ³ in juice; fully exposed berries had higher PVT	fruit exposure increased wine color and sensory ratings for linalool, rose and spice aromas no impact on wine taste, aftertaste and mouthfeel	6
	Seyval	2	shoot and cluster thinning reduced yield	little impact on TSS, TA, pH	wines issued from the cluster thinning treatment were preferred by the sensory panel only in the cooler year; under “normal” conditions, panelists rated the wines similarly	7
shoot thinning	La Crescent	1	similar cluster weight, reduced berry weight			8
training system	Kunleány	nd	single curtain reduced winter frosts			9
	Seyval	4	GDC ^b , 6-AK and YT had highest yields and lowest cluster weight	GDC and YT had lowest TSS and pH	GDC wines had lowest% ethanol and TA; GDC wines had higher intensity on melon notes, had low earthy and vegetal character, and had the lowest astringency	10
	Traminette	5	SH had highest yield compared to HC	SH and HC had higher TA, no impact on FVT ^c and PVT		11
	Traminette	2	GDC had highest berry weight compared to SH, SD and HC	VSP, SH and SD had the lowest TA; VSP had the highest juice pH; VSP and HC increased the level of FVT in juice		12
training system and cluster thinning	Louise Swenson	2	cluster thinning significantly reduced yield	TWC had higher TSS compared to VSP; cluster thinning increased TSS level in must		13

^a (1) Morris et al., 1987; (2) Barthe et al., 2012; (3) Wolpert et al., 1983; (4) Portz et al., 2010; (5) Reynolds et al., 1986 (6) Skinkis et al., 2010; (7) Berkey et al., 2011; (8) Rolfes et al., 2012; (9) Balogh, 1989; (10) Reynolds et al., 2004; (11) Bordelon et al., 2008; (12) Ji and Dami, 2008; (13) Provost et al., 2012b.

^b 6AK: 6-arms Kniffin; GDC: Geneva double curtain; HC: High cordon; SD: Smart-Dyson; SH: Scott Henry; TWC: Topwire Cordon; YT: Y-trellis.

^c FVT: free volatile terpenes; PVT: potentially-volatile terpenes.

ethyl carbamate in wine (Vincenzini et al., 2009). Studies comparing the level of biogenic amine in FRG and *V. vinifera* wines showed contradictory results (Baucom et al., 1986; Soleas et al., 1999), suggesting that large differences exist between varieties. Similarly, studies showed higher level of biogenic amines in organic compared to conventional wine (reviewed by Provost and Pedneault, 2016).

3.3.3. Tannins and color

In contrast with *V. vinifera*, FRG wines contain high levels of diglycosylated and acetylated anthocyanins and high levels of delphinidin-3-*O*-glucoside and petunidin-3-*O*-glucoside that may provide their wines with purple-blue colors (Table 4) (Manns and Mansfield, 2012; Manns et al., 2013). This trait varies largely among FRG varieties.

Color instability and lack of mouthfeel are major issues in red FRG wines (Manns et al., 2013; Springer and Sacks, 2015). FRG wines generally carry low tannin levels (≤ 200 mg/L CE eq.) and high anthocyanin levels (200–1200 mg/L M3G eq.) (Tables 4 and 5). In comparison, red *V. vinifera* wines contain between 150 and 600 mg/L CE eq. of tannins and between 200 and 400 mg/L of anthocyanin, depending on the variety and winemaking process (Casassa and Harbertson, 2014; Kilmister et al., 2014; Pedneault et al., 2014; Springer and Sacks, 2014). Indeed, studies have demonstrated that the majority of traditional extraction processes for FRG winemaking lead to high amounts of anthocyanin rather than tannins, and yield very dark wines with little mouthfeel (Table 11, Auw et al., 1996; Manns et al., 2013; Pickering and Pour Nikfardjam, 2007; Pour Nikfardjam and Pickering, 2008).

Poor tannin extractability as well as poor retention of exogenous tannins in FRG wine was recently related to high protein concentration that may contribute to precipitate tannins during winemaking (Mansfield, 2015b; Springer and Sacks, 2014). Recently, pathogenesis-related proteins have been found to interfere with tannin retention in FRG wines (Springer et al., 2016). PR-proteins expressed in reaction to biotic stresses are the major proteins in *V. vinifera* wines; they are resistant to proteases and to low pH values, making them difficult to remove or denature (Ferreira et al., 2004).

The level of PR-proteins in wine is partly related to the disease pressure in the vineyard. For instance, organic *V. vinifera* wines were found to contain higher levels of PR-proteins compared to non-organic ones, which was attributed to their higher exposure to fungi in the field (Sauvage, 2011). The impact of disease pressure and its possible modulation of PR-protein concentration in FRG varieties grown under organic management have not been studied yet.

The solution currently proposed to resolve the tannin retention issue in FRG wine is to treat the juice or the wine with bentonite in order to precipitate the proteins and then add exogenous tannins (Springer et al., 2016).

3.3.4. Aroma

The majority of studies on the aroma of FRG wines focused on the well-known “foxy compounds” that provide the “hybrid character” to *V. labrusca*-based FRG wines. Recent GC-Olfactometry/MS analyses demonstrated that most foxy compounds such as *o*-aminophenone and methyl anthranilate are not very abundant in *V. riparia* based FRG wines (Sun et al., 2011a). Similarly, no foxy compounds were reported from GC-O/MS analyses of Frontenac wines from Minnesota (Mansfield and Vickers, 2009; Table 12).

Many red FRG wines are known for their fruitiness but herbaceous notes are also reported in certain varieties such as Cabernet cortis, Prior, Regent and Frontenac, among others (Table 12; Mansfield and Vickers, 2009; Rousseau et al., 2013). Herbaceousness could relate to the presence of methoxy-pyrazine

and/or C₆ compounds such as hexanol and *cis*-3-hexenol in wine (Mansfield and Vickers, 2009; Pedneault et al., 2013a). The level of methoxy-pyrazines has been showed to decrease significantly in Frontenac berries during berry ripening (Pedneault et al., 2013a). The C₉ aldehydes nonanal and *trans,cis*-2,6-nonadienal have been shown to reach above-threshold concentrations in red FRG wines from Eastern Canada, suggesting that they could contribute to the green notes found in certain FRG wines (Slegers et al., 2015). The concentration of *trans,cis*-2,6-nonadienal has also been showed to increase in Frontenac and Marquette berries during ripening (Pedneault et al., 2013a).

3.4. Improving FRG wine quality

Historically, assays carried out to improve the quality of FRG wine were primarily meant to reduce the occurrence of foxy flavors with the use of different winemaking processes. In 1974, carbonic maceration was found to efficiently reduce foxy flavors in red Concord (*V. labrusca*) wines (Fuleki, 1974; Table 11). Carbonic maceration (CM) was initially developed to reduce oxidation reactions occurring spontaneously in grapes in order to preserve fruit flavors (Paul, 1996b). In organic wine production, restricting contact between berries and air using CM may be of particular interest because organic grapes have been shown to carry twice as much polyphenol oxidase activity compared to conventional ones (Núñez-Delgado et al., 2006).

White FRG wines such as Chardonnell, Solaris and La Crescent generally present desirable flowery notes that may be related to compounds such as C₁₃-norisoprenoids (e.g., β -damascenone) and monoterpenes (e.g., linalool) located in berry skin (Table 12; Cadwallader et al., 2009; Savits, 2014; Liu et al., 2015). In fact, extended skin maceration (24 h cold-soak and 30 h on-skin fermentation) significantly improved the intensity of floral notes in Solaris wines, but also increased green vegetable notes (Zhang et al., 2015). Conversely, short cold-soaks (3–8 h) did not improve the aroma intensity of Traminette wine (Skinkis et al., 2010).

A recent study reported for the first time the presence of 3-mercaptohexanol in the FRG variety Cayuga, at a concentration of 195 ng/L (Musumeci et al., 2015). This compound is a highly odor potent thiol (odor perception threshold: 60 ng/L) that produces a grapefruit aroma in white wine (Musumeci et al., 2015). Cayuga white is an offspring of Seyval blanc (Seyve-Villard 5–276), a variety that has often been used during the breeding of recent FRG varieties. This suggests that 3-mercaptohexanol could be present in other FRG varieties, although its presence has not yet been investigated. Based on this finding, viticulture (e.g., nitrogen status, disease control) and winemaking practices (e.g., oxidation control) that either enhance thiol production in berries, protect thiol during winemaking, or lead to thiol expression in wine could contribute to increase the occurrence of tropical aroma in FRG wines (Musumeci et al., 2015).

Blending is one of the most efficient ways to optimize the aroma of FRG wines. Indeed, most FRG varieties present a very large and rich flavor range, and many of them have complementary flavor profiles (Slegers et al., 2015). Blending may also significantly improve wine balance, especially acidity, reduce bitterness and improve wine's overall bouquet. Such richness makes FRG varieties suitable to a large range of styles that have a high potential to appeal to consumers.

4. Conclusion

The well-documented susceptibility of *V. vinifera* cultivars to major diseases such as powdery mildew, downy mildew and botrytis is a significant challenge in organic viticulture. Increasing the

Table 11

Overview of the impact of several winemaking processes on the chemistry and the sensory perception of wines made from fungus-resistant grape varieties.

Treatments	Variety	Yeast strain	Impacts on wine			ref.
			Basic chemistry	Phenolic and volatile compounds	Sensory perception	
Extraction and alcoholic fermentation processes 3 vs 8 h cold-soak at 12 °C	Traminette	Lalvin K1-V1116			no perceptible impact on wine aroma intensity	1 ^a
– direct press after crushing – whole cluster press – 6 h cold soak – 24 h cold soak – 6 or 24 h cold soak + 30 h skin fermentation *chemical deacidification used on all treatments	Solaris	Lalvin DV10TM		24 h cold soak with or without skin fermentation increased total polyphenols; extended skin contact (24 h cold soak + 30 h skin fermentation) increased the concentration of linalool, hotrienol, α-terpineol, β-damascenone and S-methyl thioacetate in wine.	extended skin contact (24 h cold soak + 30 h skin fermentation) increased the intensity of floral (Rose, Elderflower) and green vegetable descriptors.	2
- cold-soak – enzyme addition at crush - tannins addition at crush - hot press compared to skin fermentation (7 days)	Maréchal Foch	GRE		hot press increased the level of non anthonyanin phenolic (including caftaric acid, coumaric acid, catechin, epicatechin, catechin, rutin), and the level of monoglucoside and diglucoside anthocyanins; tannin addition and hot press increased tannin concentration but had no impact on their mean degree of polymerization.		3
	Corot noir	GRE		tannin addition increased tannin concentration whereas hot press decreased it, both did not impact the mean degree of polymerization.		3
- hot press - skin fermentation (7 days)	Marquette	GRE		hot press increased anthocyanin monoglucosides and tannin concentrations.		3
- immediate press for juice - immediate press for wine - hot press for juice - hot press for wine - skin fermentation (7, 13, and 21 days)	Chambourcin	Prise de mousse		hot press (juice and wine) increased color intensity (A520 nm); 13d and 21d skin fermentation decrease color intensity compared to 7d treatment; immediat press (juice & wine) increased browning (Hue); extended on-skin fermentation decreased the level of caftaric acid, coumaric acid, and procyanidin B3, and increased the level of gallic acid; hot press increased total phenol concentration.		4
-carbonic maceration (CM) at 15 or 27 °C, for 1 or 2 weeks - skin fermentation - hot press	Concord	no starter in CM; unknown starter used in other trials		carbonic maceration reduced the occurrence of bluish tones, the concentration in total phenols and decreased the concentration in methyl anthranilate.	carbonic maceration decreased the intensity of <i>V. labrusca</i> typical flavor.	5

Table 11 (Continued)

Treatments	Variety	Yeast strain	Impacts on wine			ref.
			Basic chemistry	Phenolic and volatile compounds	Sensory perception	
two experiments: 1) <i>Standard fermentation with varying percentage of whole berries</i> ; 2) <i>CM with whole clusters (2, 4, 8 days)</i> addition of OptiRed® (inactivated yeast derivative) at the beginning of skin fermentation (19 days)	Maréchal Foch	Epernay II	the use of 100% whole berries or whole clusters decreased the level of residual sugars; 8d CM decreased TA and alcohol percentage, and increased pH	100% whole berries and whole clusters decreased total phenols and color; extended CM (8d) increased total phenols.		6
	Baco noir	EC1118		OptiRed® increased the concentration of dimers procyanidins; OptiRed® increased the concentration of tyrosol and quercetin, and decreased the concentration of caffeic acid, grape reaction product, and delphinidin.	OptiRed® had no perceptible impact on wine mouthfeel.	7,8
	Maréchal Foch	EC1118		OptiRed® increased the concentration of dimer and trimers procyanidins; OptiRed® increased the concentration of caffeic acid and decreased the concentration of caftaric acid, <i>p</i> -coumaric acid, and quercetin.	OptiRed® had no perceptible impact on wine mouthfeel.	
<i>malolactic fermentation (MLF): non inoculated control compared to inoculation with the Leuconostoc oenos strains ML-34, PSU-1, and LS-5A</i>	Chancellor, DeChaunac, Maréchal Foch	Montrachet	Non-inoculated control did not completed MLF; PSU-1 and LS-5A strains completed MLF faster than ML-34; MLF increased wine pH in Chancellor (strain LS-5A only) and Maréchal Foch, decreased TA, and increased volatile acidity in all wines.		wines fermented with PSU-1 and LS-5A were preferred over ML-34 wines; all inoculated wines were preferred to the non inoculated control.	9
Post-fermentation processes and aging <i>French (Nevers; Limousin) and American oak barrels aging compared to unoaked control</i>	Seyval	n.d.		american oak increased gallic acid content of wine at a slightly higher rate than French oak; the concentration of other non-flavonoid compounds (photocatechuic, vanillic, caffeic, syringic and <i>p</i> -coumaric acids) were no affected.	perceptible changes in sensory attributes of wines compared to unoaked control were perceptible after 7 weeks of aging; by the 12th week, wines were distinguishable according to the oak type used for aging.	10

^a (1) Skinkis et al., 2010; (2) Zhang et al., 2015; (3) Manns et al., 2013; (4) Auw et al., 1996; (5) Fuleki, 1974; (6) Miller and Howell, 1989; (7) Pour Nikfardjam and Pickering, 2008; (8) Pickering and Pour Nikfardjam, 2007; (9) Giannakopoulos et al., 1984; (10) Jindra and Gallander, 1987.

use of FRG varieties would allow significant benefits for organic and conventional growers, including reducing the number of treatments per season, increasing grape yield and reducing labor costs. In addition, FRG would allow significant reduction in the use of copper-based fungicide, therefore contributing to decrease copper accumulation in vineyard soils, especially in areas under high disease pressure.

Consumer surveys have showed that wines made from FRG varieties are at least equivalent and frequently rated as superior to *V. vinifera* wines in terms of quality. Trials conducted over the last 30 years have showed that canopy management and

winemaking practices can contribute to improved FRG wine quality. Further research on these topics are needed to provide a more comprehensive understanding of the optimal cultural and winemaking practices for FRG varieties, with an emphasis on the potential side-effects from disease resistance that could interfere with the development of high quality wines (e.g., PR-proteins).

The commercialization of organic FRG wines faces a double challenge of growing grape varieties that are generally unknown to consumers and doing it under organic management. Both FRG and organic wines previously suffered from the production of bad wines that contributed to negative opinions among consumers

Table 12
Overview of volatile compounds, impact odorants and sensory descriptors in wines from 20 fungus-resistant grape varieties.

Variety ^a	Method	Analytical		Sensory	Ref.
		Volatile compounds ¹	Concentration (µg/L)	Aroma and flavors descriptors	
<i>reds</i>					
Cabernet Cortis	sensory analysis using similar descriptors for all varieties evaluated descriptive analysis			foxy, herbaceous, bell pepper, jammy, spicy	1 ^a
Chancellor				berry, currant, earthy, vegetal	2
Frontenac	descriptive analysis & GC-O/MS	ethyl isobutyrate**	0.1–0.5 mg/L	cherry, black currant, vegetal, earthy	3
		ethyl lactate	0.2–1.5 mg/L		
		ethyl 2-methylbutanoate	30–200		
		ethyl 3-methylbutanoate	40–720		
		1-hexanol	0–0.8 mg/L		
		ethyl hexanoate	1.2–3.6		
		phenethyl alcohol	0.8–2.1 mg/L		
		diethyl succinate	1.4–4.4		
		octanoic acid	0.8–5.4		
		ethyl decanoate	1.1–3.7		
Frontenac	GC-MS & odor activity values	nonanal	40		4
		<i>trans,cis</i> -2,6-nonadienal	1.26		
		β-damascenone	3.99		
		ethyl 2-methylpropanoate	281		
		ethyl hexanoate	450		
		ethyl octanoate	1128		
		nonanal	34.8		
		<i>trans,cis</i> -2,6-nonadienal	1.44		
		β-damascenone	2.25		
		ethyl 2-methylpropanoate	281		
Maréchal Foch	GC-MS & odor activity values	ethyl hexanoate	227		4
		ethyl octanoate	781		
		nonanal	52.2		
		<i>trans,cis</i> -2,6-nonadienal	1.15		
		β-damascenone	2.47		
		linalool	36.2		
		ethyl 2-methylpropanoate	254		
		ethyl butanoate	327		
		ethyl hexanoate	803		
		ethyl octanoate	2383		
Marquette	GC-MS & odor activity values	methyl anthranilate			5
		<i>trans</i> -2-hexenal			
		hexanol			
		ethyl 3-methylbutanoate			
		nonanal			
		2-phenylethanol			
		ethyl decanoate			
Prior	Sensory analysis using similar descriptors for all varieties evaluated			foxy, fusel alcohol, herbaceous, fresh fruit, jammy	1
Regent		Sensory analysis using similar descriptors for all varieties evaluated			foxy, animal, herbaceous, spicy
Sabrevois	GC-MS & odor activity values	nonanal	33.4		4
		<i>trans,cis</i> -2,6-nonadienal	1.13		
		eugenol	23.1		
		ethyl 2-methylpropanoate	283		
		ethyl octanoate	643		
St. Croix	GC-MS & odor activity values	nonanal	30		4
		<i>trans,cis</i> -2,6-nonadienal	1.19		
		eugenol	3.38		
		ethyl 2-methylpropanoate	265		
		ethyl octanoate	768		
<i>whites</i>					
Cayuga white	GC-O/MS (Charm)	2-phenylethanol β-damascenone C ₃ and C ₄ fatty acids ethyl butyrate linalool			6

Table 12 (Continued)

Variety ^a	Method	Analytical		Sensory	Ref.
		Volatile compounds ¹	Concentration (µg/L)	Aroma and flavors descriptors	
Cabernet blanc	GC-MS Sensory analysis using similar descriptors for all varieties evaluated	3-mercaptohexanol	195 ng/L	herbaceous, citrus fruit, tropical fruit	7
					1
Chardonnay	GC-O, GC-MS, and OAVs	ethyl butanoate	147–422	fruit, floral, spicy	8
		ethyl 3-methylbutanoate	ND-25.1		
		ethyl hexanoate	721–999		
		3-methyl-1-butanol	37.5–170 ppm		
		2-phenylethanol	8.94–23.4 ppm		
		β-damascenone sotolon	1.4–3.2 detected in GC-O; not quantified		
Johanniter	sensory analysis using similar descriptors for all varieties evaluated			sulfur, white fruit in syrup, dry fruit, apricot	1
La Crescent	descriptive analysis			apricot, grapefruit, lychee, pineapple, rose	9
Muscaris	sensory analysis using similar descriptors for all varieties evaluated			sulfur, fusel alcohol, citrus fruit, tropical fruit, white fruit in syrup	1
Seyval	descriptive analysis			floral, apple, earthy, melon, vegetal	2
	GC-O/MS (Charm)	β-damascenone 2-phenylethanol methyl anthranilate ethyl 2-methylbutanoate vanillin		fruity, vegetal, caramelized, pungent	10
Solaris	descriptive analysis, GC-MS	ethyl butyrate		peach/apricot, Muscat, melon, banana, strawberry	11
		propyl acetate butyl acetate 3-methylbutyl acetate hexyl acetate trans-3-hexenyl-acetate 2-phenylethyl acetate			
	descriptive analysis			fruity, citrus, tropical fruit, flowery, elderflower	12
Vidal	GC-O/MS & OAVs	ethyl 2-methylbutyrate ^{***}	14.2		13
		β-damascenone	11.3		
		decanal	13.7		
		geranyl acetone	0.26		
		ethyl hexanoate	878		
		1-octanol	9.34		
		nerol oxide	6.98		
		4-vinylguaiacol	111		
descriptive analysis			young wines: apple, tropical fruit, citrus aged wines: cooked vegetables, straw, oxidized, pungent	14	
	GC-O/MS (Charm)	β-damascenone 2-phenylethanol linalool			

^a 3 to 9 wines per varieties.

^{**} Compounds with frequency of detection higher than 75%.

^{***} Nose-perceptible odorants.

^a (1) Rousseau et al., 2013; (2) Reynolds et al., 2004; (3) Mansfield and Vickers, 2009; (4) Slegers et al., 2015; (5) Radulović et al., 2010; (6) Chisholm et al., 1994; (7) Musumeci et al., 2015 (8) Cadwallader et al., 2009; (9) Savits, 2014; (10) Andrews et al., 1990; (11) Liu et al., 2015; (12) Linden, 2014; (13) Bowen and Reynolds, 2012; (14) Chisholm et al., 1995.

concerning these products. Therefore, significant efforts are necessary to demonstrate and improve the quality of organic FRG wines, and have them accepted by consumers. According to the Web of Science database, the number of scientific publications involving FRG varieties increased by 86% between 2011 and 2016, when compared to the 2005–2010 period (search restricted to “fungus resistant grape or varieties” and “interspecific hybrid grape or varieties”). Such rise in the interest for FRG opens the door to great expectations

regarding the future of these varieties, in both organic and conventional viticulture.

Acknowledgements

The authors wish to thank Gaëlle Dubé, viticulture specialist, for valuable information on organic grape production from FRG varieties, and Dr. Ross Stevenson (UQAM), for kindly reviewing the

English writing of this article. The authors declare no conflicts of interest in the authorship or publication of this contribution.

References

- Andrews, J.T., Heymann, H., Ellersieck, M., 1990. Sensory and chemical analyses of Missouri Seyval blanc wines. *Am. J. Enol. Vitic.* 41, 116–120.
- Auw, J.M., Blanco, V., O'keefe, S.F., Sims, C.A., 1996. Effect of Cabernet sauvignon, Chambourcin, and noble wines and juices. *Am. J. Enol. Vitic.* 47, 279–286.
- Avenard, J.C., Bernos, L., Grand, O., Samir, B., 2003. Manuel de Production Intégrée en Viticulture. Éditions Féret. pp. 222.
- Balogh, I., 1989. High-trunked Horticultural University Dissertations (Hungary). pp. 167.
- Barthe, C., Dorais, M., Dubé, G., Angers, P., Pedneault, K., Impact of Cluster Thinning on Maturity and Grape Quality of Seyval blanc and Vandal-Cliche, two Hybrid Grape Varieties Grown in Quebec, Canada. *Vitinord*, Neubrandenburg, Germany, Nov. 28–Dec. 1, 2012. 10.13140/2.1.4220.7688.
- Barthe, C., 2015. Impact de la Charge Fruitière sur la Maturité et la Qualité du Raisin chez le Seyval Blanc et le Vandal Cliche, Deux Cépages Hybrides Cultivés au Québec. Master Degree Thesis. Université Laval, Québec, Canada, pp. 121.
- Baucom, T.L., Tabacchi, M.H., Cottrell, T.H.E., Richmond, B.S., 1986. Biogenic amine content of New York state wines. *J. Food Sci.* 51, 1376–1377, <http://dx.doi.org/10.1111/j.1365-2621.1986.tb13130.x>.
- Bavougian, C.M., Read, P.E., Walter-Shea, E., 2012. Training system effects on sunlight penetration, canopy structure yield, and fruit characteristics of Frontenac grapevine (*Vitis* spp.). *Int. J. Fruit Sci.* 12, 402–409, <http://dx.doi.org/10.1080/15538362.2012.679178>.
- Bavougian, C.M., Read, P.E., Schlegel, V.L., Hanford, K.J., 2013. Canopy light effects in multiple training systems on yield, soluble solids, acidity, phenol and flavonoid concentration of 'Frontenac' grapes. *HortTechnology* 23, 86–92.
- Bayramoglu, Z., Gundogmus, E., 2008. Cost efficiency on organic farming: a comparison between organic and conventional raisin-producing households in Turkey. *Span. J. Agric. Res.* 1, 3–11.
- Becker, A., 2013. Piwis in der Praxis. *Schweiz. Z. Obst Weinbau* 3, 4–7.
- Berkey, T.G., Mansfield, A.K., Lerch, S.D., Meyers, J.M., Heuvel, J.E.V., 2011. Crop load adjustment in 'Seyval Blanc' winegrape: impacts on yield components, fruit composition, consumer wine preferences, and economics of production. *HortTechnology* 21, 593–598.
- Besnard, E., Chenu, C., Robert, M., 2001. Influence of organic amendments on copper distribution among particle-size and density fractions in champagne vineyard soils. *Environ. Pollut.* 112, 329–337, [http://dx.doi.org/10.1016/S0269-7491\(00\)00151-2](http://dx.doi.org/10.1016/S0269-7491(00)00151-2).
- Bonn, M.A., Cronin, J.J., Cho, M., 2015. Do environmental sustainable practices of organic wine suppliers affect consumers' behavioral intentions? The moderating role of trust. *Cornell Hosp. Q.* 1–17, <http://dx.doi.org/10.1177/1938965515576567>.
- Bordelon, B.P., Skinkis, P.A., Howard, P.H., 2008. Impact of training system on vine performance and fruit composition of Traminette. *Am. J. Enol. Vitic.* 59, 39–46.
- Bowen, A.J., Reynolds, A.G., 2012. Odor potency of aroma compounds in riesling and vidal blanc table wines and icewines by gas chromatography-olfactometry-mass spectrometry. *J. Agric. Food Chem.* 60, 2874–2883, <http://dx.doi.org/10.1021/jf203314j>.
- Byrne, M.E., Howell, G.S., 1978. Initial response of Baco noir grapevines to pruning severity, sucker removal, and weed control. *Am. J. Enol. Vitic.* 29, 192–198.
- Cadwallader, K.R., Lopez, J.R., Menke, S.D., Surakarnkul, R., 2009. Aroma components of wines from Chardonnay: a French American hybrid grape. In: Shahidi, F. (Ed.), *Chemistry of Wine Flavor, Chemistry, Nutrition, and Health Effects*. American Chemical Society, Washington, DC, pp. 365–378, <http://dx.doi.org/10.1021/bk-2004-0871.ch027>.
- Carisse, O., Lefebvre, A., 2011. Evaluation of northern grape hybrid cultivars for their susceptibility to anthracnose caused by *Elsinoe ampelina*. *Plant Health Prog.*, <http://dx.doi.org/10.1094/PHP-2011-0805-01-RS>.
- Casassa, L.F., Harbertson, J.F., 2014. Extraction, evolution, and sensory impact of phenolic compounds during red wine maceration. *Ann. Rev. Food Sci. Technol.* 5, 83–109, <http://dx.doi.org/10.1146/annurev-food-030713-092438>.
- Chisholm, M.G., Guiher, L.A., Vonah, T.M., Beaumont, J.L., 1994. Comparison of some French-American hybrid wines with white Riesling using gas chromatography-olfactometry. *Am. J. Enol. Vitic.* 45, 201–212.
- Chisholm, M.G., Guiher, L.A., Zaczekiewicz, S.M., 1995. Aroma characteristics of aged Vidal blanc wine. *Am. J. Enol. Vitic.* 46, 56–62.
- Collective, 2008. Viticulture Biologique Presented at the Journées Techniques, Drôme, France, November 26–27 2008 (accessed on December 14 2015) <http://www.itab.asso.fr/downloads/actes%20suite/actes-viti-2008-web.pdf>.
- Dami, I., Bordelon, B., Ferree, D.C., Brown, M., Ellis, M.A., Williams, R.N., Doohan, D., 2005a. Midwest Grape Production Guide. Bulletin 919. Ohio State University Extension, pp. 156.
- Dami, I., Ferree, D.C., Kurtural, S.K., Taylor, B.H., 2005b. Influence of crop load on 'Chambourcin' yield, fruit quality, and winter hardiness under Midwestern United States environmental conditions. *Acta Hort.* 689, 203–208.
- Dami, I., Ferree, D., Prajitna, A., Scurlock, D., 2006. A five-year study on the effect of cluster thinning on yield and fruit composition of 'Chambourcin' grapevines. *HortScience* 41, 586–588.
- Daniela, P., Federica, G., Mirella, G., Diego, T., 2013. Performance of interspecific grapevine varieties in North-East Italy. *Agric. Sci.* 4, 91–101.
- Di Gaspero, G., Foria, S., 2015. Molecular Grapevine Breeding Techniques. In: *Grapevine Breeding Programs for the Wine Industry*. Elsevier, New York, pp. 23–37, <http://dx.doi.org/10.1016/B978-1-78242-075-0.00002-8>.
- Di Gaspero, G., Copetti, D., Coleman, C., Castellarin, S.D., Eibach, R., Kozma, P., et al., 2012. Selective sweep at the Rpv3 locus during grapevine breeding for downy mildew resistance. *TAG Theor. Appl. Genet.* 124, 277–286, <http://dx.doi.org/10.1007/s00122-011-1703-8>.
- Dubé, G., Turcotte, I., 2011. Guide. Richard Grenier ed. pp. 215.
- Ehrhardt, C., Arapitsas, P., Stefanini, M., Flick, G., Mattivi, F., 2014. Analysis of the phenolic composition of fungus-resistant grape varieties cultivated in Italy and Germany using UHPLC-MS/MS. *J. Mass Spectrom.* 49, 860–869, <http://dx.doi.org/10.1002/jms.3440>.
- Eibach, R., Töpfer, R., 2003. Success in resistance breeding: 'REGENT' and its steps into the market. *Acta Hort.* 603, 687–691.
- Emanuelli, F., Battilana, J., Costantini, L., Grando, M.S., 2011. Molecular breeding of grapevine for aromatic quality and other traits relevant to viticulture. In: *Breeding for Fruit Quality*. John Wiley & Sons, Inc., Hoboken, NJ, USA, pp. 247–260, <http://dx.doi.org/10.1002/9780470959350>.
- Emling, J., Sabbatini, P., 2013. Marquette Crop Load and Training System Trial for Michigan Northern Grape Project. <http://northerngrapesproject.org/wp-content/uploads/2015/02/MI-Marquette-crop-load-and-training-study.pdf>.
- Eurostat, 1999–2009. European Commission. Online Database of European Statistics (Accessed on August 15th 2015) <http://ec.europa.eu/eurostat/data/database>.
- Ferreira, R.B., Monteiro, S.S., Piçarra-Pereira, M.A., Teixeira, A.R., 2004. Engineering grapevine for increased resistance to fungal pathogens without compromising wine stability. *Trends Biotechnol.* 22, 168–173, <http://dx.doi.org/10.1016/j.tibtech.2004.02.001>.
- Flores-Vélez, L.M., Ducaroir, J., Jaunet, A.M., Robert, M., 1996. Study of the distribution of copper in an acid sandy vineyard soil by three different methods. *Eur. J. Soil Sci.* 47, 523–532, <http://dx.doi.org/10.1111/j.1365-2389.1996.tb01852.x>.
- Fragoulis, G., Trevisan, M., Di Guardo, A., Sorce, A., Van Der Meer, M., Capri, E., 2009. Development of a management tool to indicate the environmental impact of organic viticulture. *J. Environ. Qual.* 38, 826–835, <http://dx.doi.org/10.2134/jeq2008.0182>.
- Fuleki, T., Ricardo da Silva, J.M., 1997. Catechin and procyanidin composition of seeds from grape cultivars grown in Ontario. *J. Agric. Food Chem.* 45, 1156–1160, <http://dx.doi.org/10.1021/jf960493k>.
- Fuleki, T., 1974. Application of carbonic maceration to change the bouquet and flavor characteristics of red Table wines made from concord grapes. *J. Inst. Can. Sci. Technol. Aliment.* 7, 269–273, [http://dx.doi.org/10.1016/s0315-5463\(74\)73926-8](http://dx.doi.org/10.1016/s0315-5463(74)73926-8).
- Fuller, K.B., Alston, J.M., Sambucci, O.S., 2014. The value of powdery mildew resistance in grapes: evidence from California. *Wine Econ. Pol.* 3, 90–107, <http://dx.doi.org/10.1016/j.wep.2014.09.001>.
- Galbrun, C., 2008. Étude INRA: Comment Réduire ses Coûts de Production de 50%. Réussir Vigne, France (Online): <http://vigne.reussir.fr/actualites/etude-inra-comment-reduire-ses-couts-de-production-de-50-6ZKt15TA.html>, Accessed on Nov. 20 2015.
- Galet, P., 1999. Précis De Pathologie Viticole, 3ième édition. JF impression, pp. 264.
- Giannakopoulos, P.I., Markakis, P., Howell, G.S., 1984. The influence of malolactic strain on the fermentation on wine quality of three eastern red wine grape cultivars. *Am. J. Enol. Vitic.* 35, 1–4.
- Grape Growers of Ontario, 2015. 67th Annual Report (Accessed on October 25, 2015) <http://www.grapegrowersofontario.com/sites/default/files/GGO%202015%20Annual%20Report%202.pdf>.
- Guesmi, B., Serra, T., Kallas, Z., 2012. The productive efficiency of organic farming: the case of grape sector in Catalonia. *Span. J. Agric. Res.* 10, 552–566.
- Haggerty, L.L., 2013. Ripening Profile of Grape Berry Acids and Sugars in University of Minnesota Wine Grape Cultivars, Select *Vitis Vinifera*, and Other Hybrid Cultivars Master Degree Thesis. University of Minnesota, pp. 90.
- Hemstad, P.R., Luby, J.J., 2000. Utilization of *Vitis riparia* for the development of new wine varieties with resistance to disease and extreme cold. *Acta Hort.* 528, 487–490.
- Ji, T., Dami, I.E., 2008. Characterization of free flavor compounds in Traminette grape and their relationship to vineyard training system and location. *J. Food Sci.* 73, 262–267.
- Jindra, J.A., Gallander, J.F., 1987. Effect of American and French oak barrelson the phenolic composition and sensory quality of Seyval blanc wines. *Am. J. Enol. Vitic.* 38, 133–138.
- Khanizadeh, S., Rekika, D., Levasseur, A., Groleau, Y., Richer, C., Fisher, H., 2005. The effects of different cultural and environmental factors on grapevine growth, winter hardiness and performance, in three locations, in Canada. *Small Fruit Rev.* 4, 3–28.
- Khanizadeh, S., Rekika, D., Porgès, L., Levasseur, A., Groleau, Y., Fisher, H., 2008. Soluble solids acidity, canopy fruit distribution, and disease susceptibility of selected grape cultivars in Quebec. *Int. J. Fruit Sci.* 8, 200–215.
- Kilmister, R.L., Mazza, M., Baker, N.K., Faulkner, P., Downey, M.O., 2014. A role for anthocyanin in determining wine tannin concentration in Shiraz. *Food Chem.* 152, 475–482, <http://dx.doi.org/10.1016/j.foodchem.2013.12.007>.
- Komárek, M., Čadková, E., Chrástný, V., Bordas, F., Bollinger, J.C., 2010. Contamination of vineyard soils with fungicides: a review of environmental and toxicological aspects. *Environ. Int.* 36, 138–151, <http://dx.doi.org/10.1016/j.envint.2009.10.005>.

- Lee, C.Y., Robinson, W.B., Van Buren, J.P., Acree, T.E., Stoewsand, G.S., 1975. Methanol in wines in relation to processing and variety. *Am. J. Enol. Vitic.* 26, 184–187.
- Li, X., Li, L., Wang, J., Liu, Y., Guo, J., 2015. Introduction experiment of the cold resistant wine grape cultivar Frontenac. *Acta Hort.* 1082, 61–62.
- Linden, J., 2014. Sensory Profiling of Swedish White Wines and a Contextual Analysis of Swedish Viticulture Master Degree Thesis. Swedish University of Agricultural Sciences, pp. 74.
- Lisek, J., 2010. Yielding and healthiness of selected grape cultivars for processing in central Poland. *J. Fruit Orn. Plant Res.* 18, 265–272.
- Liu, J., Toldam-Andersen, T.B., Petersen, M.A., Zhang, S., Arneborg, N., Bredie, W.L.P., 2015. Instrumental and sensory characterisation of Solaris white wines in Denmark. *Food Chem.* 166, 133–142. <http://dx.doi.org/10.1016/j.foodchem.2014.05.148>.
- Mann, S., Ferjani, A., Reissig, L., 2010. What matters to consumers of organic wine? *Brit. Food J.* 114, 272–284.
- Manns, D., Mansfield, A.K., 2012. A core–shell column approach to a comprehensive high-performance liquid chromatography phenolic analysis of *Vitis vinifera* L. and interspecific hybrid grape juices, wines, and other matrices following either solid phase extraction or direct injection. *J. Chromatogr. A* 1251, 111–121. <http://dx.doi.org/10.1016/j.chroma.2012.06.045>.
- Manns, D.C., Coquard Lenerz, C.T.M., Mansfield, A.K., 2013. Impact of processing parameters on the phenolic profile of wines from hybrid red grapes Maréchal Foch, Corot noir, and Marquette. *J. Food Sci.* 78, C696–C702. <http://dx.doi.org/10.1111/1750-3841.12108>.
- Mansfield, A.K., Vickers, Z.M., 2009. Characterization of the aroma of red Frontenac table wines by descriptive analysis. *Am. J. Enol. Vitic.* 60, 430–441. <http://dx.doi.org/10.5344/ajev.2011.10067>.
- Mansfield, A.K., 2015a. Yeast Assimilable Nitrogen (YAN) Optimization for Fermentation of Cold Climate Cultivars. Project Report, Northern Grapes Project (Accessed on October 20th 2015) <http://northerngrapesproject.org/wp-content/uploads/2015/02/YAN.pdf>.
- Mansfield, A.K., 2015b. Building the Perfect Body: Tannin Strategies for Red Hybrid Wines. Northern Grapes Project Webinar Series. <https://www.youtube.com/watch?v=fEYAk0Wkgal&feature=youtu.be>.
- Martinson, T.E., Particka, C.A., 2013. Marquette Training Trial. Northern Grape Project. <http://northerngrapesproject.org/wp-content/uploads/2015/02/Marquette-Training-Trials.pdf>.
- Merdinoglu, D., Caranta, C., 2013. Quel déploiement de variétés de vignes résistantes au mildiou et à l'oïdium? In: *Les cépages résistants Aux Maladies Cryptogamiques: Panama Européen*. Groupe ICV, Bordeaux, pp. 54–59.
- Miller, D.P., Howell, G.S., 1989. The effect of various carbonic maceration treatments on must and wine composition of Maréchal Foch. *Am. J. Enol. Vitic.* 40, 170–174.
- Miller, D.P., Howell, G.S., Flore, J.A., 1996. Influence of shoot number and crop load on potted Chambourcin grapevines I. Morphology and dry matter partitioning. *Am. J. Enol. Vitic.* 47, 380–388.
- Morris, J.R., Sims, C.A., Bourque, J.E., Oakes, J.L., 1984a. Relationship of must pH and acidity to the level of soluble solids in six French-American hybrid grapes. *Ark. Farm Res.* 33, 4–5.
- Morris, J.R., Sims, C.A., Bourque, J.E., Oakes, J.L., 1984b. Influence of training system, pruning severity, and spur length on yield and quality of six French-American hybrid grape cultivars. *Am. J. Enol. Vitic.* 35, 23–27.
- Morris, J.R., Sims, C.A., Striegler, R.K., Cackler, S.D., Donley, R.A., 1987. Effects of cultivar, maturity, cluster thinning, and excessive potassium fertilization on yield and quality of Arkansas wine grapes. *Am. J. Enol. Vitic.* 38, 260–264.
- Morris, J.R., Main, G.L., Oswald, O.L., 2004. Flower cluster and shoot thinning for crop control in French-American hybrid grapes. *Am. J. Enol. Vitic.* 55, 423–426.
- Mount Kobau Wine Services, 2014, 2014. British Columbia Wine Grape Acreage Report. <http://grapegrowers.bc.ca/pdf/2014AcreageSurveyReport.pdf>.
- Musumeci, L., Ryonia, I., Pan, B., Loscos, N., Feng, H., Cleary, M., Sacks, G., 2015. Quantification of polyfunctional thiols in wine by HS-SPME-GC-MS following extractive alkylation. *Molecules* 20, 12280–12299. <http://dx.doi.org/10.3390/molecules200712280>.
- Núñez-Delgado, E., Sánchez-Ferrer, A., García-Carmona, F.F., López-Nicolás, J.M., 2006. Effect of organic farming practices on the level of latent polyphenol oxidase in grapes. *J. Food Sci.* 70, C74–C78. <http://dx.doi.org/10.1111/j.1365-2621.2005.tb09024.x>.
- Naugler, C., McCallum, J.L., Klassen, G., Strommer, J., 2007. Concentrations of trans-resveratrol and related stilbenes in Nova Scotia wines. *Am. J. Enol. Vitic.* 58, 117–119.
- Nisbet, M.A., Martinson, T.E., Mansfield, A.K., 2014. Accumulation and prediction of yeast assimilable nitrogen in New York winegrape cultivars. *Am. J. Enol. Vitic.* 65, 325–332. <http://dx.doi.org/10.5344/ajev.2014.13130>.
- Organisation Internationale de la Vigne et du Vin, 2011. Commendium of International Methods of Analysis: Maximum Acceptable Limits of Various Substances Contained in Wine (Accessed on October 20th 2015) <http://www.oiv.int/oiv/files/OIV-MA-C1-01..EN.pdf>.
- Paul, H.W., 1996a. The fall of the hybrid empire and the *Vinifera* victory. In: *Science, Vine and Wine in Modern France*. Cambridge University Press, Cambridge, pp. 99–120. <http://dx.doi.org/10.1017/cbo9780511529283.006>.
- Paul, H.W., 1996b. Languedoc-Roussillon: Innovations in Traditional Oenology. In: *Science, Vine and Wine in Modern France*. Cambridge University Press, Cambridge, pp. 260–272. <http://dx.doi.org/10.1017/CBO9780511529283.011>.
- Pavloušek, P., Kumšta, M., Matejičuková, P., 2014. Adaptation of New Resistant Grapevine Varieties to the Terroir in the Czech Republic. Congrès Internationaux des Terroirs Vitivinicols, Tokaj (Online) <http://congresdesterroirs.org/articles/voir/284>, Accessed on February 12, 2016.
- Pavloušek, P., 2010. Experiences with the cultivation characteristics of new fungus-resistant varieties for red wine production. *Mitt. Klosterneubg.* 60, 355–362.
- Pavloušek, P., 2013. Evaluation of foliar resistance of grapevine genetic resources to downy mildew (*Plasmopara viticola*). *Acta Univ. Agric. Silv. Mendel. Brun.* 60, 191–198.
- Pedneault, K., Shan Ching Seong, M., Angers, P., Determination of Quality Attributes Driving Consumer Acceptance for Cold Hardy Grape Wines Produced in Quebec. Vitinord, Neubrandenburg, Germany, November 28–December 1 2012. <http://doi.org/10.13140/2.1.1902.4326>.
- Pedneault, K., Dorais, M., Angers, P., 2013a. Flavor of cold-hardy grapes: impact of berry maturity and environmental conditions. *J. Agric. Food Chem.* 61, 10418–10438. <http://dx.doi.org/10.1021/jf402473u>.
- Pedneault, K., Slegers, A., Gagné, M.P., Angers, P., 2013b. Flavor of hybrid grapes for winemaking: a survey of the main varieties grown in Quebec (Canada) for red wine production. In: *Annual Conference of the American Journal of Enology and Viticulture-Eastern Section, Winston-Salem, USA, July 14–18th 2013* (Abstract published in the *Am. J. Enol. Vitic.* 64, 416A–424A).
- Pedneault, K., Gagné, M.P., Slegers, A., Angers, P., 2014. Phenolic and aroma composition of grapes and wines from five hybrid grape varieties used in northern wine production. In: *Annual Conference of the American Journal of Enology and Viticulture, Austin, USA, July 14–17th*.
- Pedneault, K., Montminy, G., Barthe, C., Dubé, G., Dorais, M., Angers, P., Ripening Hybrid Grapes in Cold-Climate Conditions: Results from studies conducted in Quebec, Canada 2011–2014. Vitinord, Nebraska City, USA, November 13th, 2015. (Online: <http://www.vitinord2015.org/friday-presentations.html>).
- Pickering, G.J., Pour Nikfardjam, M.S., 2007. Influence of variety and commercial yeast preparation on red wine made from autochthonous Hungarian and Canadian grapes. Part II. Oral sensations and sensory: instrumental relationships. *Eur. Food Res. Technol.* 227, 925–931. <http://dx.doi.org/10.1007/s00217-007-0807-5>.
- Plocher, T., Parke, B., 2008. *Northern Winework. Growing Grape and Making Wine in Cold Climates*, second edition. Northern Winework, pp. 213.
- Pool, R., 1995. The SARE cornell organic grape project. In: *Proceeding of the Organic Grape and Wine Production Symposium*, Geneva, NY, pp. 7–15 (Online) <https://ecommons.cornell.edu/bitstream/handle/1813/17525/1995%203rd%20Saulis%20Symposium.pdf?sequence=4&isAllowed=y>, Accessed on January 22 2016.
- Portz, D.N., Riesselman, L.B., Seeley, C., Bearer, P., Nonnecke, G.R., 2010. Effects of leaf removal on fruit quality of wine grapes grown in Iowa. *Iowa State Research Farm Progress Reports*. Paper 140.
- Pour Nikfardjam, M.S., Pickering, G.J., 2008. Influence of variety and commercial yeast preparation on red wine made from autochthonous Hungarian and Canadian grapes. Part I: phenolic composition. *Eur. Food Res. Technol.* 227, 1077–1083. <http://dx.doi.org/10.1007/s00217-008-0822-1>.
- Prajitna, A., Dami, I.E., Steiner, T.E., Ferree, D.C., Scheerens, J.C., Schwartz, S.J., 2007. Influence of cluster thinning on phenolic composition, resveratrol, and antioxidant capacity in Chambourcin wine. *Am. J. Enol. Vitic.* 58, 346–350.
- Prell, H.H., Day, P., 2001. Host plant tolerance. In: *Plant-Fungal Pathogen Interaction*. Springer Berlin Heidelberg, Berlin, Heidelberg, pp. 183–184. http://dx.doi.org/10.1007/978-3-662-04412-4_10.
- Provenzano, M.R., Bilali, E., Simeone, H., Baser, V., Mondelli, N., Cesari, D., 2010. Copper contents in grapes and wines from a Mediterranean organic vineyard. *Food Chem.* 122, 1338–1343. <http://dx.doi.org/10.1016/j.foodchem.2010.03.103>.
- Provost, C., Pedneault, K., 2016. The Organic Vineyard as a Balanced Ecosystem: Improved Organic Grape Management and its Impact on Wine Quality. *Sci. Hort.* (accepted Jan. 1st 2016).
- Provost, C., Zerouala, L., Bastien, R., D'Hauteville, J. Evaluation of the Agronomic and Oenological Characteristics of Promising Varieties in Quebec. Vitinord 2012. Neubrandenburg, Germany, November 28–December 1st 2012.
- Provost, C., D'Hauteville, J., Bastien, R., Impact of Thinning and Training Mode on the Vine and Grape Quality for Four Hybrid Varieties Grown in Quebec, Canada. Vitinord, Neubrandenburg, Germany, November 28–December 1st 2012.
- Provost, C., Bastien, R., d'Hauteville, J., 2013. Évaluation des Caractéristiques Oenologiques des Cépages Prometteurs du Québec. Final Report. Projet #6579. Programme Canadien d'Adaptation Agricole (PCAA). Centre de Recherche Agroalimentaire de Mirabel. pp. 75.
- Radulović, N., Blagojević, P., Palić, R., 2010. Volatiles of the grape hybrid cultivar othello (*Vitis vinifera* × (*Vitis labrusca* × *Vitis riparia*)) cultivated in Serbia. *J. Essent. Oil Res.* 22, 616–619.
- Ratnasooriya, C.C., Rupasinghe, H.P.V., Jamieson, A.R., 2011. Juice quality and polyphenol concentration of fresh fruits and pomace of selected Nova Scotia-grown grape cultivars. *Can. J. Plant Sci.* 90, 193–205. <http://dx.doi.org/10.4141/CJPS09137>.
- Reisch, B. I., Pool, R. M., Peterson, D. V., Martens, M. H., Henick-Kling, T., Wine and juice grape varieties for cool climates. *Information Bull.* 233. Cornell Cooperative Ext. Publ., 1993, 16 pp.
- Reynolds, A.G., Vanden Heuvel, J.E., 2009. Influence of grapevine training systems on vine growth and fruit composition: a review. *Am. J. Enol. Vitic.* 60, 251–268.
- Reynolds, A.G., Pool, R.M., Mattick, L.R., 1986. Influence of cluster exposure on fruit composition and wine quality of Seyval blanc grapes. *Vitis* 25, 85–95.

- Reynolds, A.G., Wardle, D.A., Naylor, A.P., 1995. Impact of training system and vine spacing on vine performance and berry composition of Chancellor. *Am. J. Enol. Vitic.* 46, 88–97.
- Reynolds, A.G., Wardle, D.A., Cliff, M.A., King, M., 2004. Impact of training system and vine spacing on vine performance berry composition, and wine sensory attributes of Seyval and Chancellor. *Am. J. Enol. Vitic.* 55, 84–95.
- Reynolds, A.G. (Ed.), 2015. Elsevier, Cambridge, UK, p. 425, <http://dx.doi.org/10.1016/B978-1-78242-075-0.01002-4>.
- Rolfes, D.P., Nonnecke, G.R., Domoto, P.A., 2012. Canopy Management Practices and Light Interception of Northern Grape Cultivars. Iowa State Research Farm Progress Reports. Paper 1916.
- Rolfes, D.P., 2014. The Effects of Canopy Management Practices on Fruit Quality of Northern-hardy Interspecific Hybrids of *Vitis* Spp. Master Degree Thesis. Iowa State University, Ames, USA, pp. 69.
- Rousseau, J., Chanfreau, S., Bontemps, É., 2013. Les Cépages Résistants et Maladies Cryptogamiques. Groupe ICV, Bordeaux, pp. 228.
- Salinari, F., Giosue, S., Tubiello, F.N., Rettori, A., Rossi, V., Spanna, F., Rosenweig, C., Gullino, M.L., 2006. Downy mildew (*Plasmopara viticola*) epidemics on grapevine under climate change. *Glob. Change Biol.* 12, 1299–1307, <http://dx.doi.org/10.1111/j.1365-2486.2006.01175.x>.
- Sauvage, F.-X., 2011. Vin blanc et Protéines. Presented at the Colloque IFV Sud-Ouest, MontPELLIER. <https://www.youtube.com/watch?v=v5wQOpq.d8k>.
- Savits, J.R., 2014. Descriptive Sensory Analysis of Wines Produced from Iowa-grown La Crescent Grapes Master Degree Thesis. University of Iowa, Ames, IA, pp. 99.
- Siegfried, W., Temperli, T., 2008. Piwi-Reben im Vergleich—ein Zwischenbericht. *Schweiz. Z. Obst Weinbau* 17, 6–9.
- Sivčev, B.V., Sivčev, I.L., Ranković-Vasić, Z.Z., 2010. Natural process and use of natural matters in organic viticulture. *J. Agric. Sci.* 55, 195–215.
- Skinkis, P.A., Bordelon, B.P., Butz, E.M., 2010. Effects of sunlight exposure on berry and wine monoterpenes and sensory characteristics of Traminette. *Am. J. Enol. Vitic.* 61, 147–156.
- Slegers, A., Angers, P., Ouellet, É., Truchon, T., Pedneault, K., 2015. Volatile compounds from grape skin, juice and wine from five interspecific hybrid grape cultivars grown in québec (Canada) for wine production. *Molecules* 20, 10980–11016, <http://dx.doi.org/10.3390/molecules200610980>.
- Sloan, P., Legrand, W., Krauss, K., 2010. The integration of fungus tolerant vine cultivars in the organic wine industry: the case of German wine producers. *Enomologica* 3, 37–50.
- Soleas, G.J., Carey, M., Goldberg, D.M., 1999. Method development and cultivar-related differences of nine biogenic amines in Ontario wines. *Food Chem.* 64, 49–58, [http://dx.doi.org/10.1016/S0308-8146\(98\)00092-2](http://dx.doi.org/10.1016/S0308-8146(98)00092-2).
- Springer, L.F., Sacks, G.L., 2014. Protein-precipitable tannin in wines from *Vitis vinifera* and interspecific hybrid grapes (*Vitis* spp.): differences in concentration extractability, and cell wall binding. *J. Agric. Food Chem.* 62, 7515–7523, <http://dx.doi.org/10.1021/jf5023274>.
- Springer, L.F., Sherwood, R.W., Sacks, G.L., 2016. Pathogenesis-related proteins limit the retention of condensed tannin additions to red wines. *J. Agric. Food Chem.* 9, <http://dx.doi.org/10.1021/acs.jafc.5b04906> (in press).
- Stewart, A.C., Butzke, C.E., 2012. Amino acid profiles and yeast assimilable nitrogen in hybrid winegrapes from the eastern United States. *Am. J. Enol. Vitic.* 63, 579A–589A.
- Stewart, A.C., 2013. Nitrogen Composition of Interspecific Hybrid and *Vitis Vinifera* Wine Grapes from the Eastern United States Doctoral Dissertation. Purdue University, IN, pp. 227.
- Striby, T., 2006. Évaluation intermédiaire des variétés PIWI: Propriétés agronomiques et potentiel oenologiques. In: Proceeding of the Congrès De Viticulture Biologique, Olten, Germany, March 8th 2006 (Accessed on October 20th 2006) <https://www.fibl.org/fileadmin/documents/shop/1454-viticulture.pdf>.
- Sun, Q., Gates, M.J., Lavin, E.H., Acree, T.E., Sacks, G.L., 2011a. Comparison of odor-active compounds in grapes and wines from *Vitis vinifera* and non-foxy American grape species. *J. Agric. Food Chem.* 59, 10657–10664, <http://dx.doi.org/10.1021/jf2026204>.
- Sun, Q., Sacks, G., Lerch, S., Vanden Heuvel, J.E., 2011b. Impact of shoot thinning and harvest date on yield components fruit composition, and wine quality of Maréchal Foch. *Am. J. Enol. Vitic.* 62, 32–41, <http://dx.doi.org/10.5344/ajev.2010.10023>.
- Sun, Q., Sacks, G.L., Lerch, S.D., Vanden Heuvel, J.E., 2012. Impact of shoot and cluster thinning on yield fruit composition, and wine quality of Corot noir. *Am. J. Enol. Vitic.* 63, 49–56, <http://dx.doi.org/10.5344/ajev.2011.11029>.
- Tuchschmid, A., Vonesch, G., Wins, T., 2006. Evaluation agronomique de cépages résistants aux maladies fongiques à Wädenswil, pour l'année 2005. In: Proceeding of the Congrès De Viticulture Biologique, Olten, Germany, March 8th 2006 (Accessed on October 20th 2006) <https://www.fibl.org/fileadmin/documents/shop/1454-viticulture.pdf>.
- Tuck, B., Gartner, W., 2013. Vineyards and Grapes for the North: A Status Report (Accessed on October 20th 2015) <http://www.extension.umn.edu/community/economic-impact-analysis/reports/docs/2013-Vineyards-Grapes-North.pdf>.
- Van Der Meer, M., Léville, D., 2010. Acceptation des vins de cépages résistants par les consommateurs. *Rev. Suisse Viti. Arb. Hort.* 42, 147–150.
- Verhagen, B.W.M., Trotel-Aziz, P., Couderchet, M., Hofte, M., Aziz, A., 2010. *Pseudomonas* spp.-induced systemic resistance to *Botrytis cinerea* is associated with induction and priming of defense responses in grapevine. *J. Exp. Bot.* 61, 249–260, <http://dx.doi.org/10.1093/jxb/erp295>.
- Vincenzini, M., Guerrini, S., Mangani, S., Granchi, L., 2009. Amino acid metabolisms and production of biogenic amines and ethyl carbamate. In: *Biology of Microorganisms on Grapes, in Must and in Wine*. Springer Berlin Heidelberg, Berlin, pp. 167–180, http://dx.doi.org/10.1007/978-3-540-85463-0_9.
- Vos, R., 2014. Stage of Maturation, Crop Load, and Shoot Density Affect the Fruit Quality of Cold-hardy Grape Cultivars Doctoral Dissertation. Iowa State University, Ames, IA, pp. 120.
- Weigle, T., Carroll, J., 2015. Production Guide for Organic Grapes. New York State Integrated Pest Management Program, Ithaca, NY, pp. 74.
- Wiedemann-Merdinoglu, S., Hoffmann, C., 2010. New Resistant Grape Varieties: Bottlenecks and Conditions for Adoption in Different European Grapevine-growing Regions (Online) <http://www.endure-network.eu/content/download/5586/43605/file/Grapevine%20Case%20Study%20Guide%20Number%205.pdf>, Accessed on November 30th 2015.
- Willer, H., Lernoud, J., 2015. The World of Organic Agriculture. Statistics and Emerging Trends 2015. FiBL-IFOAM Report. Research Institute of Organic Agriculture (FiBL), Frick, and IFOAM-Organics International, Bonn, 306.
- Wolf, T.K., 2008. Wine grape. In: *Production Guide for Eastern North America. Plant and Life Sciences Publishing, New York*, pp. 336.
- Wolpert, J.A., Howell, G.S., Mansfield, T.K., 1983. Sampling vidal blanc grapes. I. Effect of training system, pruning severity, shoot exposure, shoot origin, and cluster thinning on cluster weight and fruit quality. *Am. J. Enol. Vitic.* 34, 72–76.
- Wood, D.F., Looney, N.E., 1977. Some cluster thinning and gibberellic acid effects on juice and wine quality of De Chaunac grapes. *Can. J. Plant Sci.* 57, 643–646.
- Wu, J., Zhang, Yali, Lu, J., 2014. Towards understanding the mechanism of host resistance to downy mildew disease of grapevine by using genome wide expression profiling analysis. *Acta Hort.* 1046, 179–186, <http://dx.doi.org/10.17660/ActaHortic.2014.1046.23>.
- Zabadal, T.J., Vanee, G.R., Dittmer, T.W., Ledebuhr, R.L., 2002. Evaluation of strategies for pruning and crop control of Concord grapevines in southwest Michigan. *Am. J. Enol. Vitic.* 53, 204–209.
- Zhang, S., Petersen, M., Liu, J., Toldam-Andersen, T., 2015. Influence of pre-fermentation treatments on wine volatile and sensory profile of the new disease tolerant cultivar Solaris. *Molecules* 20, 21609–21625, <http://dx.doi.org/10.3390/molecules201219791>.
- Zhu, L., Zhang, Y., Deng, J., Li, H., Lu, J., 2012. Phenolic concentrations and antioxidant properties of wines made from North American grapes grown in China. *Molecules* 17, 3304–3323, <http://dx.doi.org/10.3390/molecules17033304>.
- Zini, E., Raffaeiner, M., Di Gasparo, G., Eibach, R., Grando, M.S., Letschka, T., 2015. Applying a defined set of molecular markers to improve selection of resistant grapevine accessions. *Acta Hort.* 7, 3–78, <http://dx.doi.org/10.17660/ActaHortic.2015.1082.9>.